

Exploration of *Annona muricata* (Annonaceae) in the Treatment of Hyperlipidemia Through Network Pharmacology and Molecular Docking

(Penerokaan *Annona muricata* (Annonaceae) dalam Rawatan Hiperlipidemia Melalui Rangkaian Farmakologi dan Dok Molekul)

RENY SYAHRUNI¹, ABDUL HALIM UMAR^{1,*}, HINDRIYANI NURUL RAHMAN¹ & WISNU ANANTA KUSUMA²

¹Division of Pharmaceutical Biology, College of Pharmaceutical Sciences Makassar (Sekolah Tinggi Ilmu Farmasi Makassar), Jalan Perintis Kemerdekaan Km. 13.7 Daya, Makassar 90242, Indonesia

²Department of Computer Science, Faculty of Mathematics and Natural Sciences, Jalan Meranti-Dramaga Campus, IPB University, Bogor 16680, Indonesia

Received: 11 August 2022/Accepted: 5 January 2023

ABSTRACT

Soursop (*Annona muricata* L.) is one of the plants that have antihyperlipidemic effects, but its underlying mechanism of action remains unknown. Previous investigations used TCMSP, KNApSACk, ETCM, SwissTargetPrediction, SuperPred, CTD, and TTD to identify potential targets of soursop as antihyperlipidemic. Therefore, this study aims to explore soursop active compounds and demonstrate their mechanisms against hyperlipidemia through network pharmacology and molecular docking. OB and drug-likeness properties of the compounds from *A. muricata* were screened based on Lipinski's Ro5 (Lipinski's Rule of Five) parameters. Subsequently, the network of the compound–target–disease–pathways was constructed using Cytoscape. The target PPI (protein-protein interaction) network was built using STRING and the core targets were analyzed using GO with KEGG. The main active compounds against the targets were confirmed by molecular docking analysis. Based on the results, 158 compounds were identified in *A. muricata*, and the human body was found to absorb 56. It was discovered that 20 compounds were associated with cholesterol disease. The highest degree of the disease pathway of target compounds disease was annomuricin, XDH, myocardial ischemia, and metabolic pathways, respectively. The PPI showed GAPDH (glyceraldehyde-3-phosphate dehydrogenase) protein also has the highest degree. BP, CC, MF, and KEGG enrichments that play important roles are the response to drugs, plasma membranes, protein binding, and metabolic pathways. The molecular docking experiment confirmed the correlation between ligands and receptors (quercetin-XDH, coclaurine-ADRB3, fisetin, and robinetin-XDH) with binding energies of -9.3 ; -8.9 ; and -8.8 kcal mol⁻¹, respectively. The interactions between ligands and receptors are hydrogen, alkyl, Pi-alkyl, Pi-sigma, and van der Waals bonds. It was discovered that *A. muricata* provided therapeutic effects, involving multi-compounds, multi-targets, multi-diseases, and multi-pathways as well as deep insight into the pathogenesis of hyperlipidemia. This can be used to design new drugs and develop novel therapies to treat hyperlipidemia.

Keywords: AMPK signaling; antihyperlipidemia; herbal medicine; phytochemicals; soursop

ABSTRAK

Durian belanda (*Annona muricata* L.) adalah salah satu tumbuhan yang mempunyai kesan antihiperlipidemik, tetapi mekanisme tindakan utamanya masih tidak diketahui. Penyelidikan sebelumnya menggunakan TCMSP, KNApSACk, ETCM, SwissTargetPrediction, SuperPred, CTD dan TTD untuk mengenal pasti sasaran potensi durian belanda sebagai antihiperlipidemik. Oleh itu, kajian ini bertujuan untuk meneroka sebatian aktif durian belanda dan menunjukkan mekanismenya terhadap hiperlipidemia melalui rangkaian farmakologi dan dok molekul. OB dan sifat keserupaan dadah bagi sebatian daripada *A. muricata* telah disaring berdasarkan parameter Lipinski's Ro5 (Lipinski's Rule of Five). Selepas itu, rangkaian sebatian-sasaran-penyakit-laluan dibina menggunakan Cytoscape. Rangkaian sasaran PPI (interaksi protein-protein) dibina menggunakan STRING dan sasaran teras dianalisis menggunakan GO dengan KEGG. Sebatian aktif utama terhadap sasaran telah disahkan oleh analisis dok molekul. Berdasarkan keputusan, 158 sebatian dikenal pasti dalam *A. muricata* dan tubuh manusia didapati menyerap 56. Didapati bahawa 20 sebatian dikaitkan dengan penyakit kolesterol. Peratusan tertinggi laluan penyakit, penyakit sebatian sasaran

masing-masing ialah annomuricin, XDH, iskemia miokardium dan laluan metabolik. PPI menunjukkan protein GAPDH (glyceraldehyde-3-phosphate dehydrogenase) juga mempunyai peratusan tertinggi. Pengayaan BP, CC, MF dan KEEG yang memainkan peranan penting ialah tindak balas terhadap ubat, membran plasma, pengikatan protein dan laluan metabolik. Uji kaji dok molekul mengesahkan korelasi antara ligan dan reseptor (quercetin-XDH, coclaurine-ADRB3, fisetin, dan robinetin-XDH) dengan tenaga mengikat masing-masing -9.3 ; -8.9 ; dan -8.8 kcal mol⁻¹. *A. muricata* memberikan kesan terapeutik, melibatkan pelbagai sebatian, pelbagai sasaran, pelbagai penyakit dan pelbagai laluan serta pandangan mendalam tentang patogenesis hiperlipidemia. Ini boleh digunakan untuk mereka bentuk ubat baharu dan membangunkan terapi baru untuk merawat hiperlipidemia.

Kata kunci: Antihiperlipidemia; durian belanda; fitokimia; pengisyratan AMPK; ubat herba

INTRODUCTION

Hyperlipidemia is a risk factor for atherosclerosis, cardiovascular disease, stroke, and diabetes mellitus (Jiang et al. 2022; Nunes & Quintão 2022). It increases one or more cholesterols, phospholipids, or triglycerides, characterized by an elevation in total cholesterol, LDL (low-density lipoprotein), and TG (triglycerides) accompanied by a decrease in HDL (high-density lipoprotein) (Jing et al. 2022; Severino et al. 2020; Yao, Li & Zeng 2020). Hyperlipidemia can be treated with antihyperlipidemic drugs, but the use of synthetic drugs has more significant side effects than traditional ones (Obika et al. 2022).

In Indonesia, people in certain regions traditionally use soursop (*Annona muricata* L.) as an antihyperlipidemic. It was discovered that the roots, leaves, stem bark, and fruit pulp from plant can be used as antidiabetic and antioxidant (Agu & Okolie 2017; Handayani et al. 2022), anticancer (Gavamukulya et al. 2014), antiobesity, and alternative treatment for type 2 diabetes mellitus (Wang et al. 2021a). Furthermore, anti-inflammatory, smooth muscle relaxant, sedative, hypotensive, and antispasmodic effects are reported from the plant's leaves, barks, and roots (Moghadamtousi et al. 2015). Secondary metabolites that are contained in the leaves of *A. muricata* include alkaloids, terpenoids, saponins, and flavonoids. The previous study by Coria-Téllez et al. (2018) identified that alkaloid compounds, flavonoids, and tannins can reduce LDL cholesterol. The mechanism of active compounds in *A. muricata* such as pharmacological activities, interactions between compounds and target proteins, as well as disease pathways from the perspective of network pharmacology and molecular docking, is still unclear, specifically for antihyperlipidemic activity. Network analysis, bioinformatics, and molecular docking can be used to study the mechanism (Lu et al. 2022). For traditional

medicine, several investigations showed that network pharmacology used in conjunction with molecular docking and simulation can effectively predict pharmacological mechanisms through multi-component and multi-target interactions (Fan et al. 2022; Liu et al. 2022). Network pharmacology analysis usually uses machine learning, graph mining, molecular docking, simulation modeling, and other techniques to analyze the relationships between compounds, target proteins, diseases, and pathways. These techniques were widely carried out in the search for active compounds from plant that have the potential for antihyperlipidemic (Wang et al. 2022a).

This study used network pharmacology to explore the underlying mechanisms and targets of the *A. muricata* compound for treating hyperlipidemia. Molecular docking was also used to validate correlations between the active compounds and potential target genes. The flowchart of network pharmacology and molecular docking technology employed is shown in Figure S1.

MATERIALS AND METHODS

DATA AND TOOLS

The main active components of *A. muricata* were mined from a literature review (Abdul Wahab et al. 2018; Gavamukulya et al. 2014; Gleye et al. 2000; Naik & Sellappan 2020; Rojas-Armas et al. 2022). The scientific name of herbs was used as the keyword to retrieve all compounds, while the secondary metabolites and types were determined. The duplication of compounds was eliminated during the construction of the database. Furthermore, online databases, namely KNApSACk Family (http://www.knapsackfamily.com/KNApSACk_Family/), TCMSP (traditional Chinese medicine systems pharmacology database and analysis platform) (<http://lsp.nwsuaf.edu.cn/tcmsp.php>) (Ru et al. 2014), and ETCM (The encyclopedia of traditional Chinese medicine)

(<http://www.nrc.ac.cn:9090/ETCM/index.PHP/home/index/>), were used. The active compounds were confirmed by the online databases, namely PubChem (<https://pubchem.ncbi.nlm.nih.gov/>), ChemSpider (<http://www.chemspider.com/>), Molbase (www.molbase.com), and SpectraBase (<https://spectrabase.com/>). The molecular structure of each active compound was confirmed using the online database and stored in *.pdb and SMILES formats. The chemical structures of the compounds were converted to pdb and SMILES format using MarvinSketch 19.6 (<https://chemaxon.com/products/marvin/>) when the format was unavailable in the online database.

CALCULATION AND PREDICTION OF ABSORPTION PARAMETERS

Lipinski's Ro5 was used for predicting absorption parameters, with MW less than 500, the number of hydrogen bond donors, namely hydrogen atoms attached to O and N, nOHNH was less than 5. Furthermore, the number of hydrogen bond acceptors, which were O and N, nON, were less than 10, and the lipid water partition coefficient (miLogP) was less than 5 (Lipinski 2004). Files from the online database and MarvinSketch 19.6 in SMILES format were analyzed using Molinspiration Cheminformatics (<https://www.molinspiration.com/>) and *.pdb formats using Lipinski Ro5 (<http://www.scfbio-iitd.res.in/>). The compounds that met the parameters were selected for further analysis.

SCREENING AND PREDICTION OF POTENTIAL TARGETS

For each compound, the potential targets were predicted using SwissTargetPrediction (<http://www.swisstargetprediction.ch/>) (Gfeller et al. 2014) and SuperPred website (<http://prediction.charite.de/>) by setting the attribute of *Homo sapiens*. By removing duplicate information, the Swiss Target and SuperPred prediction were used to predict *A. muricata* target proteins. The selected targets were applied to further analysis by referring to the TTD (therapeutic target database) (<https://db.idrblab.org/ttd/>) (Li et al. 2018) to obtain target protein-related diseases.

SCREENING AND PREDICTION OF THE DISEASE PATHWAY

Disease targets were analyzed using the CTD (comparative toxicogenomics database) (<http://ctdbase.org/>) disease database. The identified diseases were integrated into the KEGG (Kyoto encyclopedia of genes and genomes) (<https://www.genome.jp/kegg/>) to obtain the most

common disease pathways associated with the target genes. Subsequently, the target was identified with a p-value of 0.01 (Davis et al. 2019) to obtain the disease and its pathways.

NETWORK PHARMACOLOGY CONSTRUCTION

Cytoscape ver. 3.8.2 (<https://cytoscape.org/>) software was used to construct the models, namely 1. multi-compounds–multi-targets, 2. multi-targets–multi-diseases, 3. multi-diseases–multi-pathways, 4. multi-compounds–targets–diseases–pathways, and 5. multi-compounds–diabetes gene targets–hyperlipidemia disease–hyperlipidemia pathways, and Merge were used to combine the two network models into the component–target–disease–pathway (C–T–D–P). The network analyzer function in the software was used to further analyze the network model's relations. Topology features were applied to evaluate network interactions, namely degree, betweenness, and closeness centrality. The nodes in different shapes and colors of the network represented active compounds, targets, diseases, and pathways, while the edges indicated their interaction. The degree refers to the number of nodes directly connected. The high degree values reflect the node's importance in the network, indicating that the node is more important. Furthermore, the active core compounds were screened, and the targets showing above-average values among the three parameters were selected based on degree values (Ye, Li & Hu 2021).

CONSTRUCTION OF PROTEIN-PROTEIN INTERACTION (PPI) NETWORK

The PPI network was constructed by importing the candidate genes to the STRING, which is the search tool for the retrieval of interacting genes, ver. 11.0 (<https://string-db.org>) database (Szklarczyk et al. 2018), with the highest confidence of 0.9. It was applied to predict potential interactions and evaluate the overlap between target genes using a unique *Homo sapiens* database. In this study, the confidence score was employed to measure the likelihood of PPI. The higher the confidence score the protein sequence pair has, the more likely the pair will interact. The results obtained were analyzed using Cytoscape ver. 3.8.2 (Saito et al. 2012) and the PPI that underlies the mechanism of hyperlipidemia was obtained. The features were used to evaluate network interactions, namely degree, betweenness, and closeness centrality. The three parameters' highest value was used to determine the target protein.

FUNCTIONAL ENRICHMENT ANALYSIS OF GO (GENE ONTOLOGY)

GO was analyzed using DAVID Bioinformatics Resources 6.8 server (<https://david.ncifcrf.gov/home.jsp>) (Sherman et al. 2022) and STRING ver. 11.0 (<https://stringdb.org>). SRplot (<http://www.bioinformatics.com.cn/en>) was used to process the data and visualize the enrichment results of GO, including BP (biological process), MF (molecular function), CC (cellular component), and KEGG.

COMPONENT-TARGET MOLECULAR DOCKING

The analysis of molecular binding between the ligand and receptor was carried out to examine the binding mode and free energy, the type and distance of bonds, as well as amino acid residues. Subsequently, two-dimensional (2D) structures of ligands were extracted from the PubChem data bank (<https://pubchem.ncbi.nlm.nih.gov/>), and the receptors that match the target (specific to *Homo sapiens*) were obtained from RCSB PDB (protein data bank) (<https://www.rcsb.org/>). This analysis used UCSF Chimera ver. 1.15 (<https://www.cgl.ucsf.edu/chimera/>) for ligand optimization (force field MMFF94 and algorithm Steepest Descent) and PyRx (<https://pyrx.sourceforge.io/>) equipped with AutoDock Vina to perform molecular docking. The conformation with the best affinity and lowest binding energy scores was selected as the final docking conformation and visualized by PyMOL ver. 2.3 (<https://pymol.org/2/>) and BIOVIA Discovery Studio 2019 (<https://www.3dsbiovia.com>). The hydrogen bonds and their binding sites were observed and analyzed. Finally, ligand-receptor interactions were displayed as 2D and 3D diagrams (Umar et al. 2022).

RESULTS

COLLECTION AND SCREENING OF CANDIDATE ACTIVE COMPOUNDS IN *A. muricata*

A total of 158 compounds were identified from *A. muricata*, while the main active components that qualified the absorption parameters by following Lipinski's Ro5 were 56 compounds, as shown in Table S1.

TARGET PREDICTION

The compounds that met the parameters of Lipinski's Ro5 were further analyzed with SwissTargetPrediction. The results obtained were 560 targets, which were examined using the Therapeutic Target Database program to identify diseases related to the targets of

each compound. The results were analyzed using CTD to determine the cholesterol disease pathway.

NETWORK PHARMACOLOGY OF *A. muricata*

A pharmacological network was created to determine the relationship between compounds, targets, diseases, and pathways of *A. muricata* compounds that met the absorption parameters, as shown in Figure 1. The nodes and edges formed in the C–T–D–P component tissue were 425 and 1056, respectively. The target compounds and disease pathways with the highest grade values in the tissue were morin, fisetin, coronin, DRD2, asthma, and cytokine-cytokine receptor interactions, as presented in Tables S2-S5.

A pharmacological network devoted to cholesterol in form of a compound–target–disease–pathway in Figure 2 caused nodes and edges of 39 and 72, respectively. Meanwhile, only 20 of the 56 compounds in *A. muricata* were associated with cholesterol disease in this network. The pharmacological network of cholesterol disease with the highest degree of compound, target, disease, and pathway were anomurine, (+)-anomurine, anomurine and coclaurine, XDH (xanthine dehydrogenase), myocardial ischemia, and metabolic pathways, respectively.

The compounds anomurine, (+)-anomurine, anomurine, and coclaurine target two cholesterol diseases. The target proteins of the four compounds are ADRB2 (adrenergic receptor beta-2) and ADRB3 (adrenergic receptor beta-3), while the disease is based on the pharmacological network, namely hyperlipidemia and myocardial ischemia. One of the pathways for hyperlipidemia is the AMPK signaling pathway (5' AMP-activated protein kinase), which involves genes that play a role in hyperlipidemia, as shown in Figure 3. The core targets that correlate with hyperlipidemia include ADRA1A, PP2A, HMGR, IGF1R, P13K, and PPAR γ .

PROTEIN-PROTEIN INTERACTION (PPI) NETWORK

Figure 4 shows that the PPI network consists of 164 nodes and 464 edges. The results of the interaction network analysis of GAPDH protein (glyceraldehyde-3-phosphate dehydrogenase) had the highest degree, betweenness, and closeness centrality of 63, 0.0721, and 0.5776, respectively, as illustrated in TABLE S6.

GO AND KEGG ENRICHMENT ANALYZE

The analysis of GO on target genes used parameters for BP, CC, MF, and KEGG disease pathways, and was

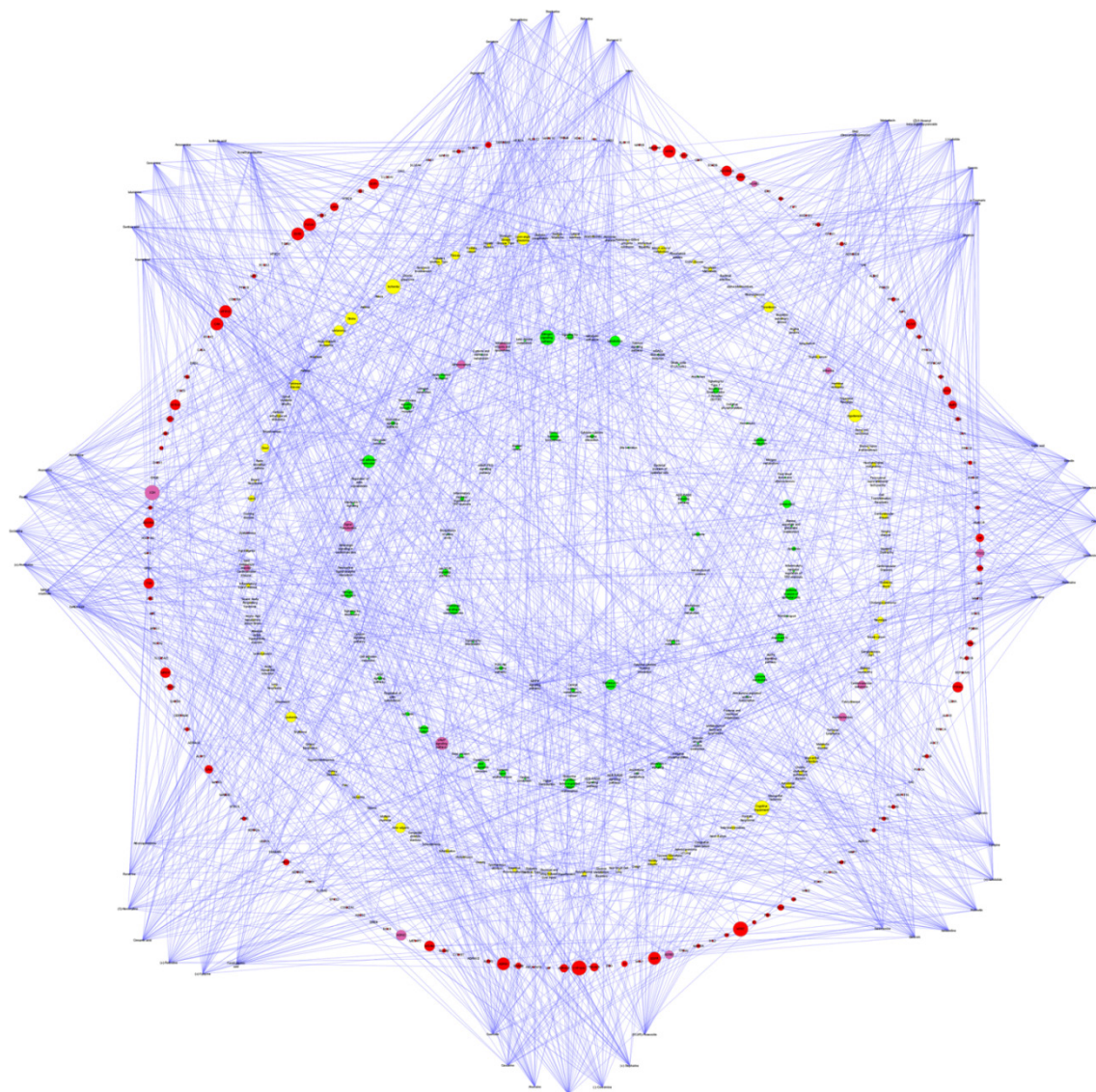


FIGURE 1. Network pharmacology of multi-compounds–multi-targets–multi-diseases–multi-pathways (C–T–D–P). The larger nodes represent more important hub nodes. Nodes in the outer ring represent the compounds. Node (inner ring) color indicates target (red), disease (yellow), and pathway (green). The purple nodes indicate each specific target, disease, and pathway associated with hyperlipidemia

evaluated with the Bonferroni method and p -value < 0.01 . Figure 5 shows the results of the top 15 GO (BP, CC, and MF) and top 25 (KEGG) in $-\log P$ value, as well as the green color in the KEGG pathway, which indicates a more significant enrichment of GO terms. Furthermore, BP, CC, MF, and the KEGG played the most roles, namely response to drugs, plasma membrane, protein binding, and metabolic pathway, with enrichment scores of 18.50, 57.10, 63.10, and 35, respectively.

RESULTS OF COMPOUNDS-TARGET MOLECULE DOCKING

Compounds and targets of pharmacological networks associated with cholesterol disease were analyzed using molecular docking. During this process, four ligands with the lowest bond affinity (negative value) were obtained, namely quercetin, coclaurine, fisetin, and robinetin with values -9.3 , -8.9 , -8.8 , and -8.8 kcal mol $^{-1}$, respectively. The residues and amino acid bonds that occur in the

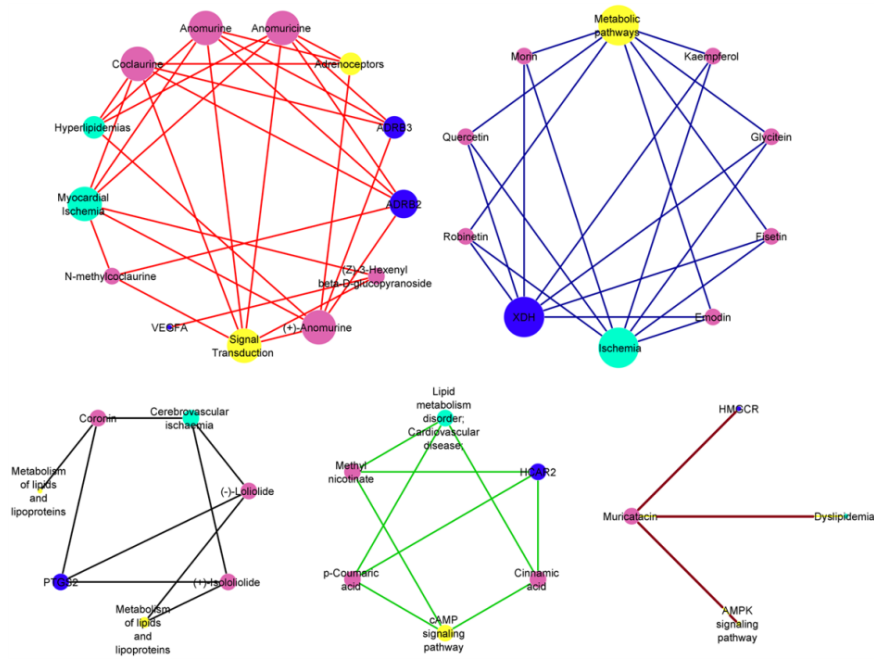


FIGURE 2. Networks between compounds-target-disease-pathways for hyperlipidemia. The larger the circle size, the more interaction there is between nodes. Node color indicates pink (compound), blue (target), green (disease), and yellow (pathway)

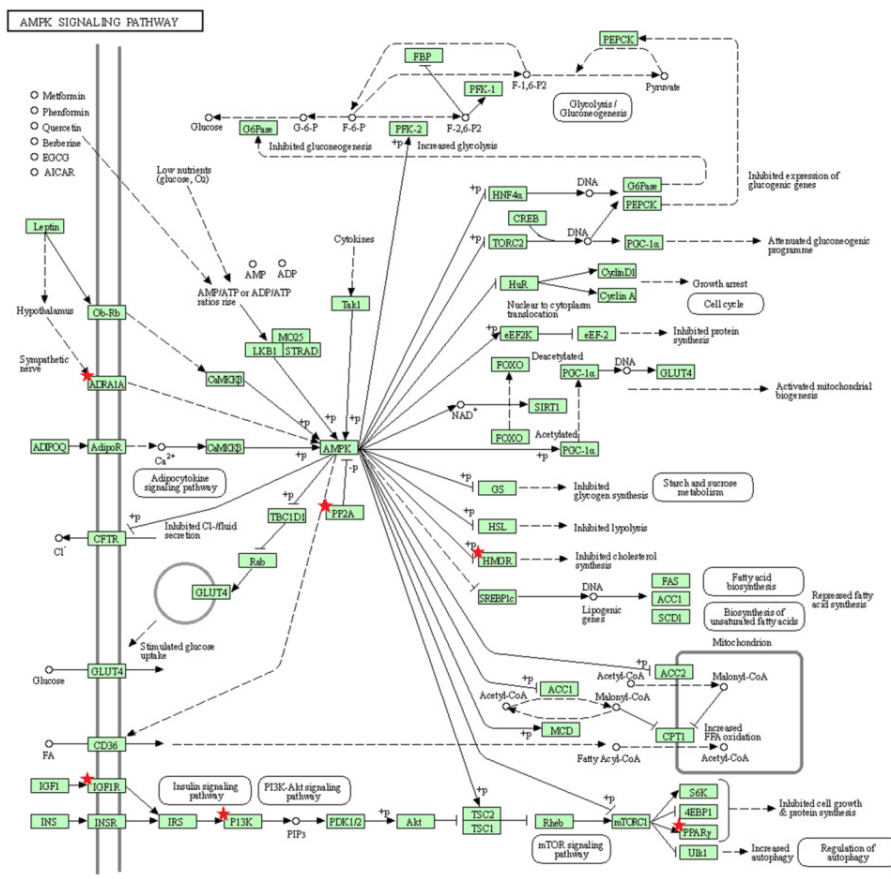


FIGURE 3. AMPK signaling pathway mechanism in hyperlipidemia. Red asterisks represent core targets

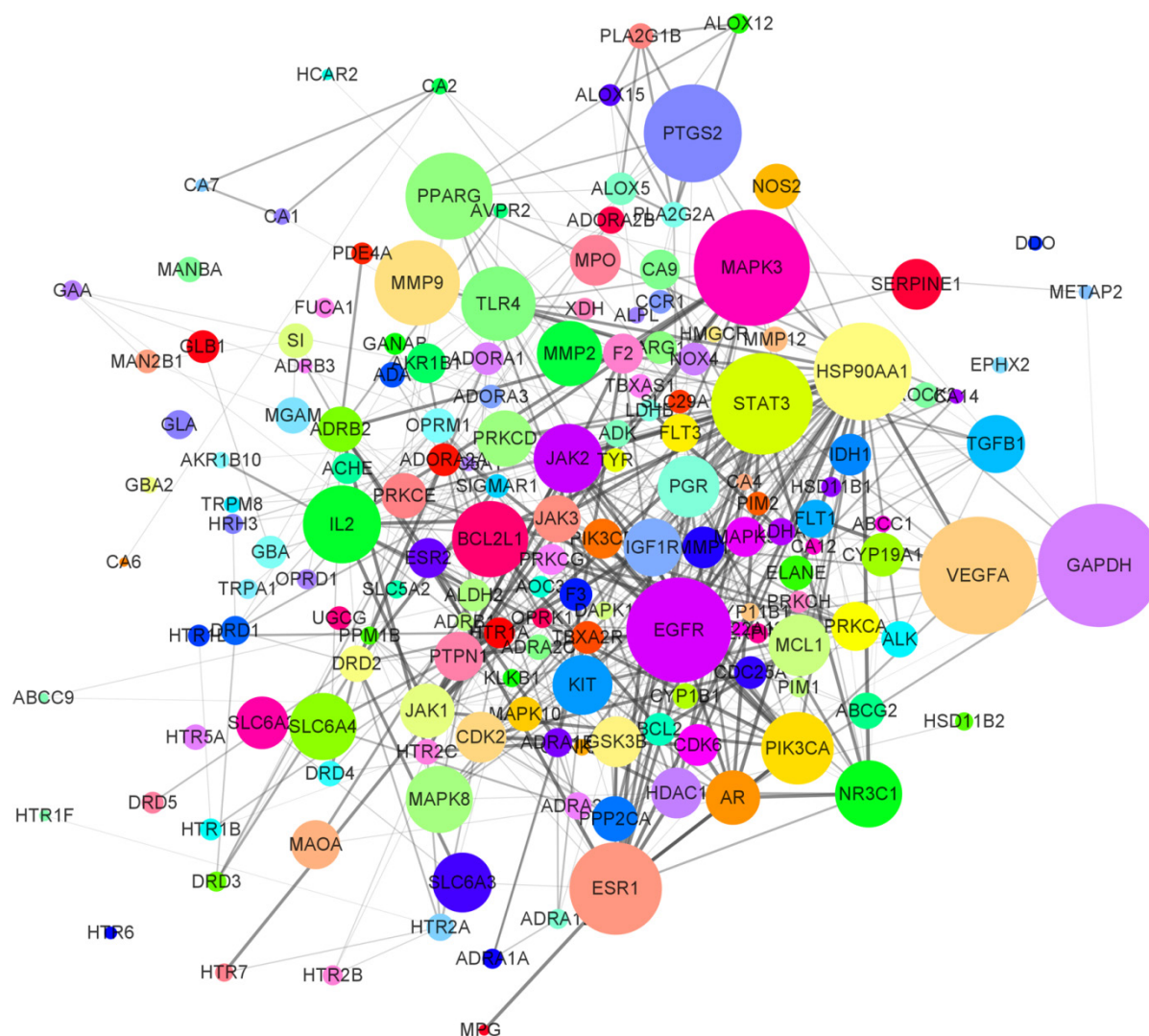


FIGURE 4. Network of protein-protein interaction. The size of the circle indicates the number of proteins correlated with the target

interaction between the four ligands and their receptors are summarized in Figure 6.

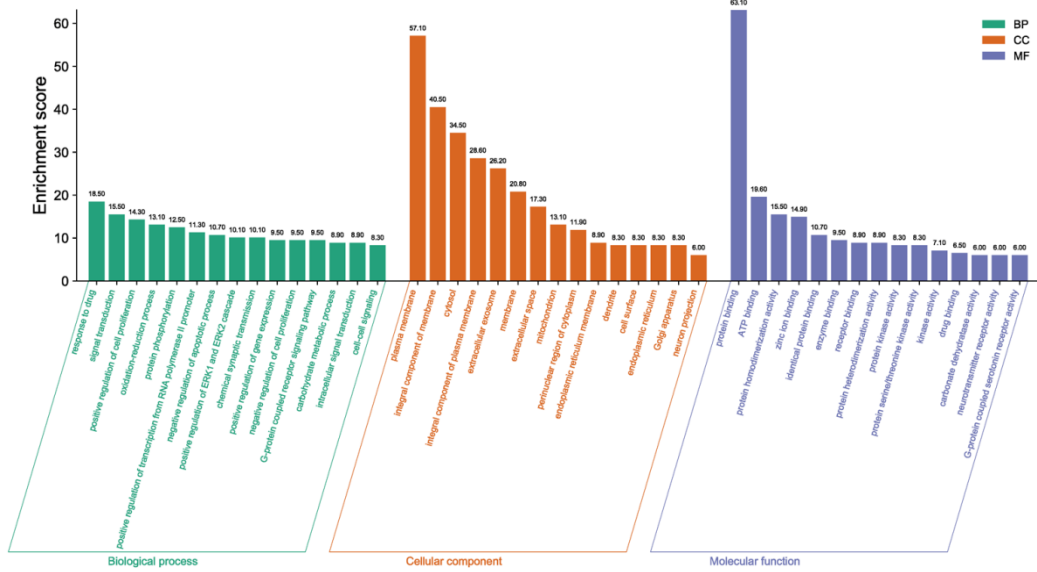
Quercetin-XDH binds through hydrogen bonds with Ser289, coclaurine-ADRB3 through Tyr59, Arg150, and Leu318, fisetin-XDH through Ile326 and Met329, as well as robinetin-XDH via Leu228, Glu295, and Ile326. The results of molecular docking, including residue, binding energy, and bond distance of the four ligands and their receptors are summarized in Table S7, and the 2D and 3D views in Figure S2.

DISCUSSION

The combination of network pharmacology and molecular

docking has been applied in this study. Both methods can be used to examine the mechanism of action of *A. muricata* compounds in treating hyperlipidemia. Exploration of herbal medicines for the management of complex diseases such as hyperlipidemia using network pharmacology is a well-accepted approach.

The development of new drug candidates is often constrained by their lack of ADME (absorption, distribution, metabolism, and excretion) properties. This high-cost nature makes the drug development process more difficult. Although ADME is an essential process in drug discovery and development, OB (oral bioavailability) is the most significant pharmacokinetic



KEGG Pathway analysis

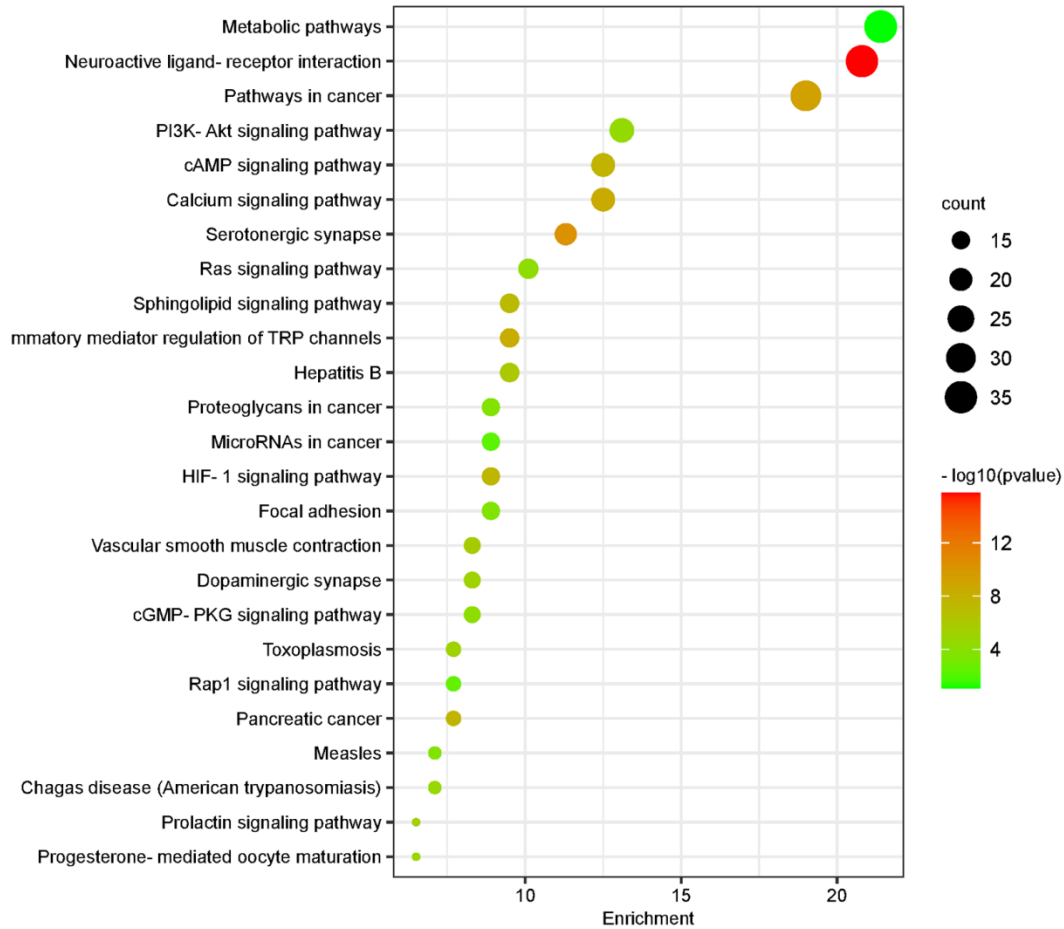


FIGURE 5. Enrichments analysis of the candidate target of *A. muricata* by gene ontology (GO) against hyperlipidemia. Biological process (BP) (green chart), cellular component (CC) (brown chart), molecular function (MF) (blue chart), KEGG pathway (bubble chart)

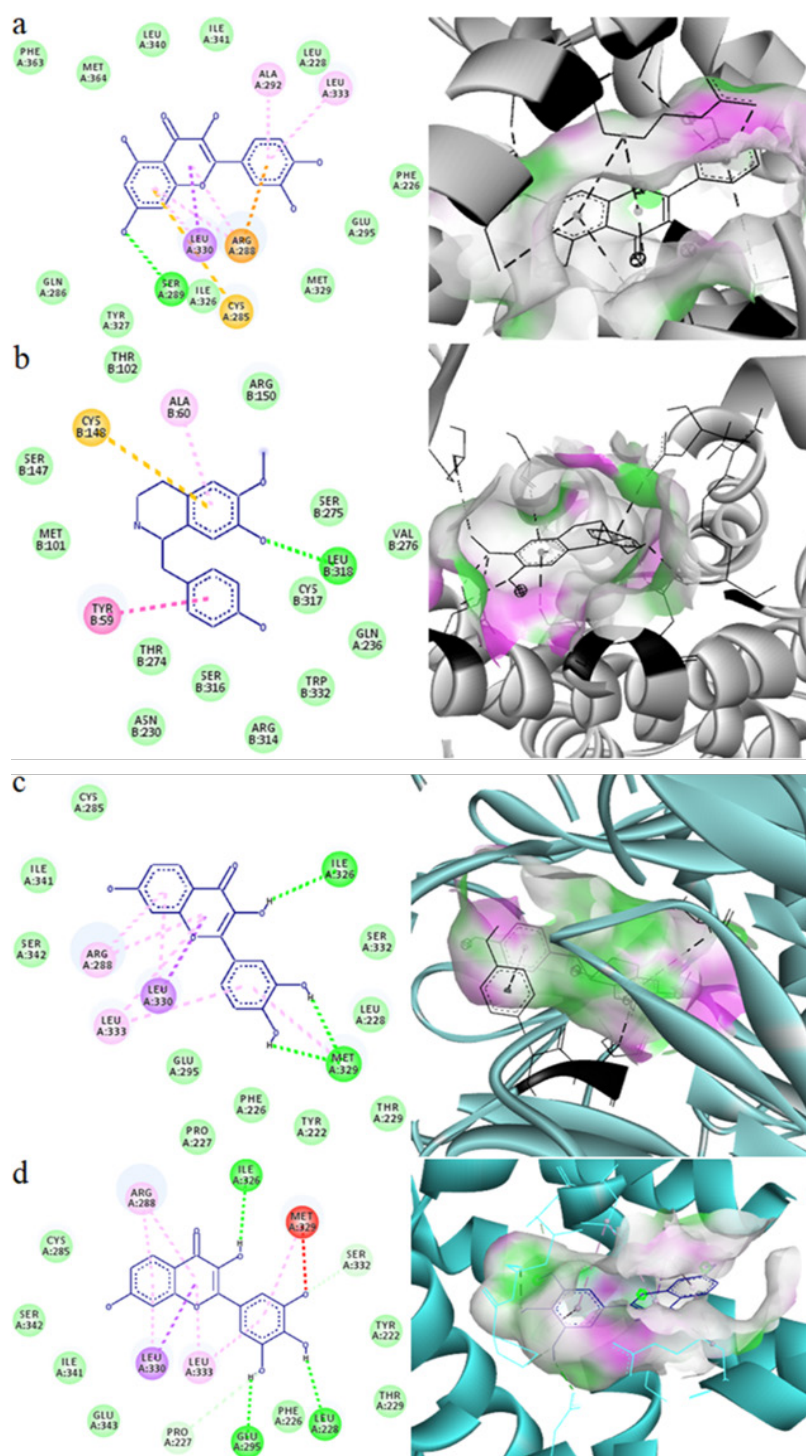


FIGURE 6. The represented results for the action mode of active compounds with target protein using molecular docking. Ligands and receptors on the protein's active site (2D and 3D view). Quercetin with target XDH (a), coclaurine and ADRB3 (b), fisetin (c) and robinetin (d) with XDH

parameter. This study identified 158 compounds from *A. muricata*, but only 56 were considered eligible. Some compounds with poor solubility and permeability cause absorption problems (Lipinski 2004). One of the parameters of an excellent medicinal compound for oral use is the ability to be adequately absorbed by the body. Furthermore, Lipinski's Ro5 parameters such as MW (molecular weight) (500), miLog P (5), nON (hydrogen bond acceptors) (5), and NOHNH (hydrogen donors) (10) can be used to determine the absorption of natural compounds (Ibrahim et al. 2021).

The results of the pharmacological network analysis showed that the compounds with the highest degree belonged to the alkaloid group, namely anomuricine, (+)-anomurine, anomurine, and coclaurine. This compound belongs to the class of alkaloids (Abdul Wahab et al. 2018; Coria-Télez et al. 2018; Pieme et al. 2014; Ragasa et al. 2012). A previous report showed that alkaloids have cholesterol-lowering activity with a mechanism of action for inhibiting dietary cholesterol uptake and HMG-CoA (hydroxy-3-methyl-glutaryl coenzyme A) reductase activity (Maharjan et al. 2022).

Alkaloid compounds interact with ADRB3 and ADRB2, with G-protein coupled receptors as the target. ADRB3 induces WAT (white adipose tissue) lipolysis and increases BAT's (brown adipose tissue) thermogenesis (Pinto, Festuccia & Magdalon 2022). ADRB3 mutations in white adipose cells can slow lipolysis, causing lipid retention in adiposity and significantly increasing leptin and dyslipidemia (Daghestani et al. 2018). ADRB2 promotes the mobilization of stored triglycerides and ADIPOQ (adiponectin), secreted by adipose tissue stimulates autophagy by inhibiting mTORC1 and promoting the activation of AMPK (Tripathi, Srivastava & Tripathi 2018). Moreover, AMPK signaling plays a crucial role in regulating cellular energy metabolism as shown in Figure 3. It is an essential metabolic sensor that regulates the body's energy balance (van der Vaart, Boon & Houtkooper 2021). An AMPK signaling molecule is highly correlated with the activation of cholesterol signaling-related molecules such as HMGCR (3-hydroxy-3-methylglutaryl-CoA reductase), FDFT1 (farnesyl-diphosphate farnesyltransferase 1), and LSS (lanosterol synthase) (Xu et al. 2022) PI3K (phosphoinositide 3-kinases) (Li et al. 2022a), PP2A (protein phosphatase 2A) (Ke et al. 2022), IGF1R (insulin-like growth factor 1 receptor) (Wang et al. 2022b), and PPAR (peroxisome proliferator-activated receptor) (Berghoff, Spieth & Saher 2022).

Based on the PPI network, GAPDH has the highest degree, betweenness, and closeness centrality, indicating that the protein has the largest correlation to target PPI (Wang et al. 2022b). GAPDH is a glycolytic enzyme that catalyzes glyceraldehyde 3-phosphate to 1,3-diphosphoglycerate and increases glycolysis, where glucose is converted into pyruvic acid. Increased ROS (reactive oxygen species) and ischemia (atherosclerosis) by GAPDH can cause damage to mitochondria (Qvit et al. 2016). Ischemia often occurs due to atherosclerosis, a condition where cholesterol plaques form on the walls of arteries due to risk factors for diseases such as hypercholesterolemia, diabetes, and hypertension (Jebari-Benslaiman et al. 2022).

Response to the drug (BP), plasma membrane (CC), and protein binding (MF) had the highest scores on GO analysis. The three GO are most likely to underlie the antihyperlipidemic mechanism of soursop active compounds. Furthermore, drug response has a relationship that can affect the therapeutic effect. The reaction catalyzed by human HMG-CoA reductase targets antihyperlipidemic drugs (statins) to lower serum cholesterol levels. HMG-CoA reductase inhibitors are agents with healthy blood cholesterol and lipid-lowering effects that are used to prevent primary and secondary atherosclerosis. The enzyme HMG-CoA reductase catalyzes the conversion of HMG-CoA to mevalonate. This four-electron oxidoreduction is a rate-limiting step in synthesizing cholesterol and other isoprenoids (Li et al. 2022b). The plasma membrane also facilitates intracellular cholesterol transport, controls cellular cholesterol balance, and influences transcriptional regulation (Islam, Hlushchenko & Pfisterer 2022). Since protein binding is associated with biological responses, changes can cause a significant difference in biological responses. According to Ghaleb et al. (2022), binding proteins on the endoplasmic reticulum cause downregulation of LDL receptors, leading to hypercholesterolemia. Metabolic pathways are related to lipid metabolism and hypercholesterolemia diseases such as triglycerides, fatty acids, cholesterol esters, and phospholipids (Wang et al. 2021b).

Jin et al. (2021) stated that the lower the affinity (bond energy), the higher the bond strength between the ligand and the target protein. This indicated that the compound has a more stable bound to the receptor, while quercetin, coclaurine, fisetin, and robinetin bind to receptors through hydrogen bonds. The formation of hydrogen bonds minimizes the energy of the complex

between the molecule-receptor, thereby making it stable (Yu et al. 2021).

The mechanism of quercetin is inhibiting low-density lipoprotein oxidation by suppressing myeloperoxidase-catalyzed oxidation and plays an anti-inflammatory, antioxidant, and lipid-lowering role in AFLD by decreasing serum and hepatic NF- κ B, p65, COX-2, and IL-6. The decreased activated AMPK (Lin et al. 2022) which contributes to the pathogenesis of liver steatosis. Lipophagy-related Rab7 has been presumed as a crucial regulator in the progression of alcohol liver disease despite elusive mechanisms. More importantly, whether or not hepatoprotective quercetin targets Rab7-associated lipophagy disorder is unknown. Herein, alcoholic fatty liver induced by chronic-plus-single-binge ethanol feeding to male C57BL/6J mice was manifested by hampering autophagosomes formation with lipid droplets and fusion with lysosomes compared with the normal control, which was normalized partially by quercetin. The GST-RILP pulldown assay of Rab7 indicated an improved GTP-Rab7 as the quercetin treatment for ethanol-feeding mice. HepG2 cells transfected with CYP2E1 showed similar lipophagy dysfunction when exposed to ethanol, which was blocked when cells were transfected with siRNA-Rab7 in advance. Ethanol-induced steatosis and autophagic flux disruption were aggravated by the Rab7-specific inhibitor CID1067700 while alleviated by transfecting with the Rab7Wt plasmid, which was visualized by immunofluorescence co-localization analysis and mCherry-GFP-LC3 transfection. Furthermore, TBC1D5, a Rab GTPase-activating protein for the subsequent normal circulation of Rab7, was downregulated after alcohol administration but regained by quercetin. Rab7 circulation retarded by ethanol and corrected by quercetin was further shown by fluorescence recovery after photobleaching (FRAP, involved the PI3K signaling pathway (Wang et al. 2022b)). Several studies showed that quercetin exerts anti-adipogenic effects in 3 T3-L1 cells by inhibiting the adipocyte-specific transcription factors PPAR- γ as well as C/EBP α and activating the AMPK signaling pathways (Chang, Lee & Chiang 2012). Coclaurine exhibited significant effects in hyperlipidemia (Yang et al. 2022), while Fisetin, a bioflavonoid possesses anti-inflammatory, antilipidemic, and anticancer properties. It was discovered that fisetin can also ameliorate and reduce oxLDL-induced upregulation of SREBP-1, thereby expressing its downstream lipid synthesis genes HMGCR and FAS, which inhibit ROS induced NLRP3 inflammasome activation

(Varghese et al. 2021). Robinetin correlated with high antioxidant activity (Sari et al. 2021), known free radicals, specifically ROS, which is another critical mechanism involved in atherosclerosis development and progression (Mollazadeh et al. 2018).

CONCLUSIONS

In this study, 56 compounds of *A. muricata* met the criteria for drug absorption parameters and pharmacological tissue, while 20 were associated with cholesterol disease. Coclaurine, anomuricine, (+)-anomurine, and anomurine compounds have the most targets, with ADRB2 and ADRB3 target proteins in hyperlipidemia and myocardial ischemia. BP, CC, MF, and KEEG have the most significant roles, namely response to drugs, plasma membranes, protein binding, and metabolic pathways. The molecular bonds of quercetin with XDH (3osi), coclaurine with ADRB3 (7dh5), fisetin, and robinetin with XDH (3osi) have binding energies of -9.3 , -8.9 , and -8.8 kcal mol $^{-1}$, respectively. This shows a holistic view of the potential pharmacological and molecular mechanisms of *A. muricata* as an antihyperlipidemic agent. The results provided new methods for investigating other traditional medicine compounds. However, limitations of this study include the use of computerized methods. Secondly, with the combination of network pharmacology and molecular docking, the therapeutic mechanism of active compounds from *A. muricata* is not fully understood. A comprehensive understanding depends on the expected development of multi-disciplines. Thirdly, the results need to be further verified by experiments.

REFERENCES

- Abdul Wahab, S.M., Jantan, I., Haque, Md.A. & Arshad, L. 2018. Exploring the leaves of *Annona muricata* L. as a source of potential anti-inflammatory and anticancer agents. *Frontiers in Pharmacology* 9: 661.
- Agu, K.C. & Okolie, P.N. 2017. Proximate composition, phytochemical analysis, and *in vitro* antioxidant potentials of extracts of *Annona muricata* (Soursop). *Food Science & Nutrition* 5(5): 1029-1036.
- Berghoff, S.A., Spieth, L. & Saher, G. 2022. Local cholesterol metabolism orchestrates remyelination. *Trends in Neurosciences* 45(4): 272-283.
- Cárdenas, C., Torres-Vargas, J.A., Cárdenas-Valdivia, A., Jurado, N., Quesada, A.R., García-Caballero, M., Martínez-Poveda, B. & Medina, M.Á. 2021. Non-targeted metabolomics characterization of *Annona muricata* leaf extracts with anti-angiogenic activity. *Biomedicine & Pharmacotherapy* 144: 112263.

- Chang, Y.C., Lee, T.S. & Chiang, A.N. 2012. Quercetin enhances ABCA1 expression and cholesterol efflux through a p38-dependent pathway in macrophages. *Journal of Lipid Research* 53(9): 1840-1850.
- Coria-Téllez, A.V., Montalvo-González, E., Yahia, E.M. & Obledo-Vázquez, E.N. 2018. *Annona muricata*: A comprehensive review on its traditional medicinal uses, phytochemicals, pharmacological activities, mechanisms of action and toxicity. *Arabian Journal of Chemistry* 11(5): 662-691.
- Daghestani, M., Daghestani, M., Daghistani, M., Eldali, A., Hassan, Z.K., Elamin, M.H. & Warsy, A. 2018. ADRB3 polymorphism rs4994 (Trp64Arg) associates significantly with bodyweight elevation and dyslipidaemias in Saudis but not rs1801253 (Arg389Gly) polymorphism in ARDB1. *Lipids in Health and Disease* 17(1): 58.
- Davis, A.P., Grondin, C.J., Johnson, R.J., Sciaky, D., McMorran, R., Wieggers, J., Wieggers, T.C. & Mattingly, C.J. 2019. The comparative toxicogenomics database: Update 2019. *Nucleic Acids Research* 47(D1): D948-D954.
- Fan, L., Feng, S., Wang, T., Ding, X., An, X., Wang, Z., Zhou, K., Wang, M., Zhai, X. & Li, Y. 2022. Chemical composition and therapeutic mechanism of Xuanbai Chengqi Decoction in the treatment of COVID-19 by network pharmacology, molecular docking and molecular dynamic analysis. *Molecular Diversity* 27(1): 81-102.
- Gavamukulya, Y., Abou-Ellella, F., Wamunyokoli, F. & AEL-Shemy, H. 2014. Phytochemical screening, anti-oxidant activity and *in vitro* anticancer potential of ethanolic and water leaves extracts of *Annona muricata* (Graviola). *Asian Pacific Journal of Tropical Medicine* 7: S355-S363.
- Gfeller, D., Grosdidier, A., Wirth, M., Daina, A., Michielin, O. & Zoete, V. 2014. SwissTargetPrediction: A web server for target prediction of bioactive small molecules. *Nucleic Acids Research* 42(W1): W32-W38.
- Ghaleb, Y., Elbitar, S., Philippi, A., El Khoury, P., Azar, Y., Andrianirina, M., Loste, A., Abou-Khalil, Y., Nicolas, G., Le Borgne, M., Moulin, P., Di-Filippo, M., Charrière, S., Farnier, M., Yelnick, C., Carreau, V., Ferrières, J., Lecerf, J.M., Derksen, A., Bernard, G., Gauthier, M.S., Coulombe, B., Lütjohann, D., Fin, B., Boland, A., Olaso, R., Deleuze, J.F., Rabès, J.P., Boileau, C., Abifadel, M. & Varret, M. 2022. Whole exome/genome sequencing joint analysis of a family with oligogenic familial hypercholesterolemia. *Metabolites* 12(3): 262.
- Gleye, C., Raynaud, S., Fourneau, C., Laurens, A., Laprévote, O., Serani, L., Fournet, A. & Hocquemiller, R. 2000. Cohibins C and D, two important metabolites in the biogenesis of acetogenins from *Annona muricata* and *Annona nutans*. *Journal of Natural Products* 63(9): 1192-1196.
- Handayani, S.I., Sari, M.I.P., Sardjana, M.S., Kusmardi, K., Nurbaya, S., Rosmalena, R., Sinaga, E. & Prasasty, V.D. 2022. Ameliorative effects of *Annona muricata* leaf ethanol extract on renal morphology of alloxan-induced mice. *Applied Sciences* 12(18): 9141.
- Ibrahim, M.A.A., Abdeljawaad, K.A.A., Abdelrahman, A.H.M. & Hegazy, M.E.F. 2021. Natural-like products as potential SARS-CoV-2 Mpro inhibitors: *in-silico* drug discovery. *Journal of Biomolecular Structure and Dynamics* 39(15): 5722-5734.
- Islam, M.M., Hlushchenko, I. & Pfisterer, S.G. 2022. Low-density lipoprotein internalization, degradation and receptor recycling along membrane contact sites. *Frontiers in Cell and Developmental Biology* 24(10): 826379.
- Jebari-Benslaiman, S., Galicia-García, U., Larrea-Sebal, A., Olaetxea, J.R., Alloza, I., Vandenbroeck, K., Benito-Vicente, A. & Martín, C. 2022. Pathophysiology of atherosclerosis. *International Journal of Molecular Sciences* 23(6): 3346.
- Jiang, L., Xiong, Y., Tu, Y., Zhang, W., Zhang, Q., Nie, P., Yan, X., Liu, H., Liu, R. & Xu, G. 2022. Elucidation of the transport mechanism of puerarin and gastrodin and their interaction on the absorption in a Caco-2 cell monolayer model. *Molecules* 27(4): 1230.
- Jin, J., Chen, B., Zhan, X., Zhou, Z., Liu, H. & Dong, Y. 2021. Network pharmacology and molecular docking study on the mechanism of colorectal cancer treatment using Xiao-Chai-Hu-Tang. *PLoS ONE* 16(6): e0252508.
- Jing, Y.S., Ma, Y.F., Pan, F.B., Li, M.S., Zheng, Y.G., Wu, L.F. & Zhang, D.S. 2022. An insight into antihyperlipidemic effects of polysaccharides from natural resources. *Molecules* 27(6): 1903.
- Ke, W., Zhou, Y., Lai, Y., Long, S., Fang, L. & Xiao, S. 2022. Porcine reproductive and respiratory syndrome virus nsp4 positively regulates cellular cholesterol to inhibit type I interferon production. *Redox Biology* 49: 102207.
- Li, D., Zhang, J. & Liu, Q. 2022a. Brain cell type-specific cholesterol metabolism and implications for learning and memory. *Trends in Neurosciences* 45(5): 401-414.
- Li, J., Zhu, F., Xu, W. & Che, P. 2022b. Therapeutic properties of isoliquiritigenin with molecular modeling studies: investigation of anti-pancreatic acinar cell tumor and HMG-CoA reductase inhibitor activity for treatment of hypercholesterolemia. *Archives of Medical Science*. <https://doi.org/10.5114/aoms/145448>
- Li, Y.H., Yu, C.Y., Li, X.X., Zhang, P., Tang, J., Yang, Q., Fu, T., Zhang, X., Cui, X., Tu, G., Zhang, Y., Li, S., Yang, F., Sun, Q., Qin, C., Zeng, X., Chen, Z., Chen, Y.Z. & Zhu, F. 2018. Therapeutic target database update 2018: Enriched resource for facilitating bench-to-clinic research of targeted therapeutics. *Nucleic Acids Research* 46(Database issue): D1121-D1127.
- Lin, H., Guo, X., Liu, J., Liu, P., Mei, G., Li, H., Li, D., Chen, H., Chen, L., Zhao, Y., Jiang, C., Yu, Y., Liu, W. & Yao, P. 2022. Improving lipophagy by restoring Rab7 cycle: Protective effects of quercetin on ethanol-induced liver steatosis. *Nutrients* 14(3): 658.
- Lipinski, C.A. 2004. Lead and drug like compounds: The rule of five revolution. *Drug Discovery Today Technologies* 1(4): 337-341.

- Liu, C., Fan, F., Zhong, L., Su, J., Zhang, Y. & Tu, Y. 2022. Elucidating the material basis and potential mechanisms of Ershiwuwei Lvxue Pill acting on rheumatoid arthritis by UPLC-Q-TOF/MS and network pharmacology. *PLoS ONE* 17(2): e0262469.
- Lu, S., Tang, L., Zhou, L., Lai, Y., Liu, L. & Duan, Y. 2022. Study on the multitarget mechanism and active compounds of essential oil from *Artemisia argyi* treating pressure injuries based on network pharmacology. *Evidence-Based Complementary and Alternative Medicine* 2022: e1019289.
- Maharjan, B., Payne, D.T., Ferrarese, I., Giovanna Lupo, M., Kumar Shrestha, L., Hill, J.P., Ariga, K., Rossi, I., Sharan Shrestha, S., Panighel, G., Lal (Swagat) Shrestha, R., Sut, S., Ferri, N. & Dall'Acqua, S. 2022. Evaluation of the effects of natural isoquinoline alkaloids on low density lipoprotein receptor (LDLR) and proprotein convertase subtilisin/kexin type 9 (PCSK9) in hepatocytes, as new potential hypocholesterolemic agents. *Bioorganic Chemistry* 121: 105686.
- Moghadamtousi, S.Z., Fadaeinasab, M., Nikzad, S., Mohan, G., Ali, H.M. & Kadir, H.A. 2015. *Annona muricata* (Annonaceae): A review of its traditional uses, isolated acetogenins and biological activities. *International Journal of Molecular Sciences* 16(7): 15625-15658.
- Mollazadeh, H., Carbone, F., Montecucco, F., Pirro, M. & Sahebkar, A. 2018. Oxidative burden in familial hypercholesterolemia. *Journal of Cellular Physiology* 233(8): 5716-5725.
- Naik, A.V. & Sellappan, K. 2020. Chromatographic fingerprint of essential oils in plant organs of *Annona muricata* L. (Annonaceae) using HPTLC. *Analytical Chemistry Letters* 10(2): 214-226.
- Nunes, V.S., da Silva Ferreira, G. & Quintão, E.C.R. 2022. Cholesterol metabolism in aging simultaneously altered in liver and nervous system. *Aging* 14(3): 1549-1561.
- Obika, P., Beamon, J., Ali, S., Kakar, N., Analla, A., Mouddeen, R.E., Shihadeh, L., Patel, S., Hudson, B., Khan, F., Puglisi-Weening, M., Basist, P., Ahmad, S. & Shahid, M. 2022. 6 - Herbal medicines for the treatment of metabolic syndrome. In: *Herbal Medicines*, edited by Sarwat, M. & Siddique, H. Massachusetts: Academic Press. hlm. 139-191.
- Pieme, C.A., Kumar, S.G., Dongmo, M.S., Moukette, B.M., Boyoum, F.F., Ngogang, J.Y. & Saxena, A.K. 2014. Antiproliferative activity and induction of apoptosis by *Annona muricata* (Annonaceae) extract on human cancer cells. *BMC Complementary and Alternative Medicine* 14(1): 516.
- Pinto, Y.O., Festuccia, W.T.L. & Magdalon, J. 2022. The involvement of the adrenergic nervous system in activating human brown adipose tissue and browning. *Hormones* 21(2): 195-208.
- Qvit, N., Joshi, A.U., Cunningham, A.D., Ferreira, J.C.B. & Mochly-Rosen, D. 2016. Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) protein-protein interaction inhibitor reveals a non-catalytic role for GAPDH oligomerization in cell death. *Journal of Biological Chemistry* 291(26): 13608-13621.
- Ragasa, C.Y., Soriano, G., Torres, O.B., Don, M.J. & Shen, C.C. 2012. Acetogenins from *Annona muricata*. *Pharmacognosy Journal* 4(32): 32-37.
- Rojas-Armas, J.P., Arroyo-Acevedo, J.L., Palomino-Pacheco, M., Ortiz-Sánchez, J.M., Calva, J., Justil-Guerrero, H.J., Castro-Luna, A., Ramos-Cevallos, N., Cieza-Macedo, E.C. & Herrera-Calderon, O. 2022. Phytochemical constituents and ameliorative effect of the essential oil from *Annona muricata* L. leaves in a murine model of breast cancer. *Molecules* 27(6): 1818.
- Ru, J., Li, P., Wang, J., Zhou, W., Li, B., Huang, C., Li, P., Guo, Z., Tao, W., Yang, Y., Xu, X., Li, Y., Wang, Y. & Yang, L. 2014. TCMSP: A database of systems pharmacology for drug discovery from herbal medicines. *Journal of Cheminformatics* 6(1): 13.
- Saito, R., Smoot, M.E., Ono, K., Ruschinski, J., Wang, P.L., Lotia, S., Pico, A.R., Bader, G.D. & Ideker, T. 2012. A travel guide to Cytoscape plugins. *Nature Methods* 9(11): 1069-1076.
- Sari, R.K., Prayogo, Y.H., Sari, R.A.L., Asidah, N., Rafi, M., Wientarsih, I. & Darmawan, W. 2021. *Intsia bijuga* heartwood extract and its phytosome as tyrosinase inhibitor, antioxidant, and sun protector. *Forests* 12(12): 1792.
- Severino, P., D'Amato, A., Pucci, M., Infusino, F., Adamo, F., Birtolo, L.I., Netti, L., Montefusco, G., Chimenti, C., Lavalle, C., Maestrini, V., Mancone, M., Chilian, W.M. & Fedele, F. 2020. Ischemic heart disease pathophysiology paradigms overview: From plaque activation to microvascular dysfunction. *International Journal of Molecular Sciences* 21(21): 8118.
- Sherman, B.T., Hao, M., Qiu, J., Jiao, X., Baseler, M.W., Lane, H.C., Imamichi, T. & Chang, W. 2022. DAVID: A web server for functional enrichment analysis and functional annotation of gene lists (2021 update). *Nucleic Acids Research* 50(W1): W216-W221.
- Szklarczyk, D., Gable, A., Lyon, D., Junge, A., Wyder, S., Huerta-Cepas, J., Simonovic, M., Doncheva, N., Morris, J., Bork, P., Jensen, L. & von Mering, C. 2018. STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. *Nucleic Acids Research* 47(D1): D607-D613.
- Tripathi, S., Srivastava, S. & Tripathi, Y.B. 2018. Obesity and its complications: Role of autophagy. *International Journal of Pharmaceutical Sciences and Research* 9(8): 3100-3113.
- Umar, A.H., Ratnadewi, D., Rafi, M., Sulistyaningsih, Y.C., Hamim, H. & Kusuma, W.A. 2022. Drug candidates and potential targets of *Curculigo* spp. compounds for treating diabetes mellitus based on network pharmacology, molecular docking and molecular dynamics simulation. *Journal of Biomolecular Structure and Dynamics*. doi: 10.1080/07391102.2022.2135597
- van der Vaart, J.I., Boon, M.R. & Houtkooper, R.H. 2021. The role of AMPK signaling in brown adipose tissue activation. *Cells* 10(5): 1122.

- Varghese, J.F., Patel, R., Singh, M. & Yadav, U.C.S. 2021. Fisetin prevents oxidized low-density lipoprotein-induced macrophage foam cell formation. *Journal of Cardiovascular Pharmacology* 78(5): e729-e737.
- Wang, B., Wang, H., Li, Y. & Song, L. 2022a. Lipid metabolism within the bone micro-environment is closely associated with bone metabolism in physiological and pathophysiological stages. *Lipids in Health and Disease* 21(1): 5.
- Wang, H., Wang, H., Zhang, J., Luo, J., Peng, C., Tong, X. & Chen, X. 2022b. Molecular mechanism of Crataegi Folium and Alisma Rhizoma in the treatment of dyslipidemia based on network pharmacology and molecular docking. *Evidence-Based Complementary and Alternative Medicine* 2022: e4891370.
- Wang, F.X., Zhu, N., Zhou, F. & Lin, D.X. 2021a. Natural aporphine alkaloids with potential to impact metabolic syndrome. *Molecules* 26(20): 6117.
- Wang, Q., Du, L., Hong, J., Chen, Z., Liu, H., Li, S., Xiao, X. & Yan, S. 2021b. Molecular mechanism underlying the hypolipidemic effect of Shanmei Capsule based on network pharmacology and molecular docking. *Technology and Health Care* 29(S1): 239-256.
- Xu, X.Y., Choi, H.S., Park, S.Y., Kim, J.K., Seo, K.H., Kim, H. & Kim, Y.J. 2022. *Hibiscus syriacus* L. cultivated in callus culture exerts cytotoxicity in colorectal cancer via Notch signaling-mediated cholesterol biosynthesis suppression. *Phytomedicine* 95: 153870.
- Yang, J., Zhang, Y., Jiang, L., Li, C., Sun, Z., Zhang, Y., Lin, T., Jiang, Y. & Liu, B. 2022. A triple combination strategy of UHPLC-MSn, hypolipidemic activity and transcriptome sequencing to unveil the hypolipidemic mechanism of *Nelumbo nucifera* alkaloids. *Journal of Ethnopharmacology* 282: 114608.
- Yao, Y.S., Li, T.D. & Zeng, Z.H. 2020. Mechanisms underlying direct actions of hyperlipidemia on myocardium: An updated review. *Lipids in Health and Disease* 19(1): 23.
- Ye, J., Li, L. & Hu, Z. 2021. Exploring the molecular mechanism of action of Yinchen Wuling powder for the treatment of hyperlipidemia, using network pharmacology, molecular docking, and molecular dynamics simulation. *BioMed Research International* 2021: e9965906.
- Yu, J., Yuan, H., Bao, L. & Si, L. 2021. Interaction between piperine and genes associated with sciatica and its mechanism based on molecular docking technology and network pharmacology. *Molecular Diversity* 25(1): 233-248.

*Corresponding author; email: ahuhalim76@yahoo.com

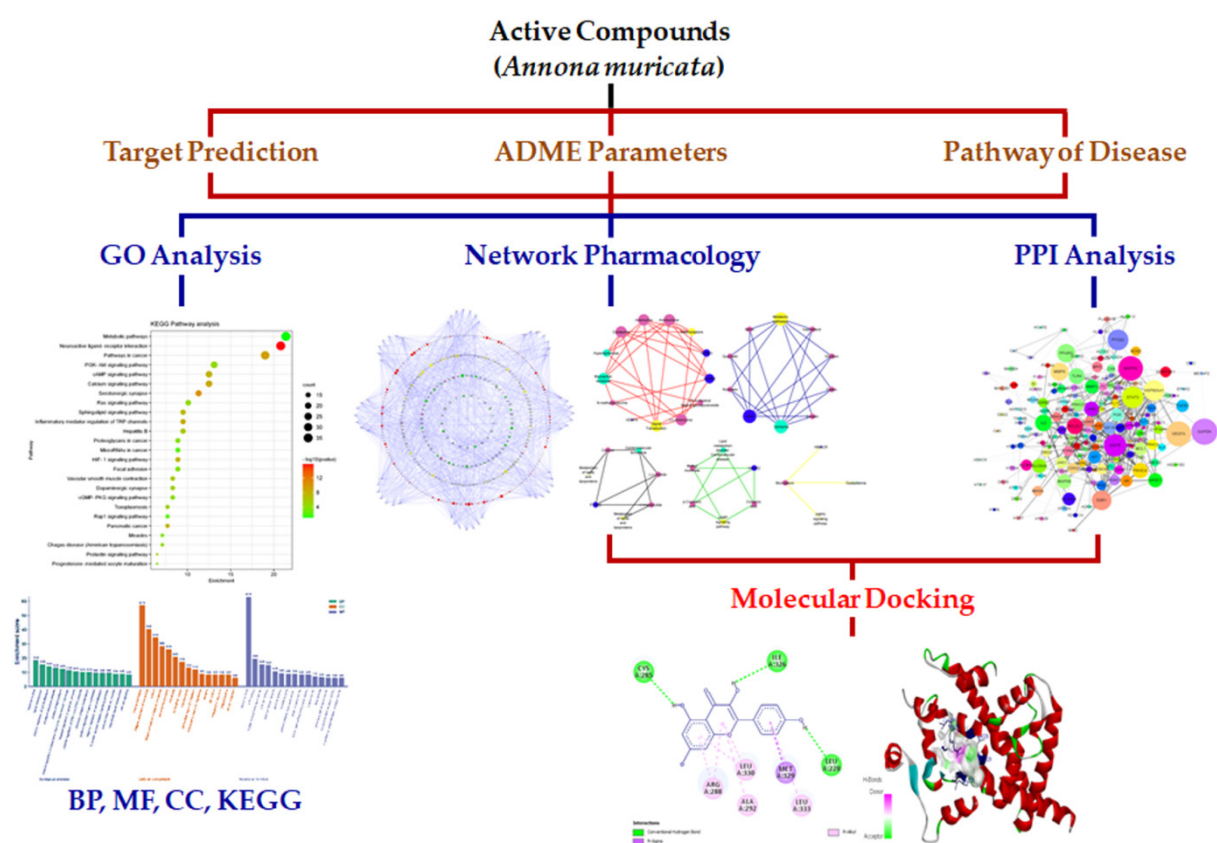


FIGURE S1. Workflow for network pharmacology and molecular docking technology

TABLE S1. *A. muricata* compound and absorption parameters

No	Compound	Absorption parameters			MW (≤ 500)	Result
		miLogP (≤ 5)	nOHNH (≤ 5)	nON (≤ 10)		
1	p-Coumaric acid	1.43	2	3	164.16	✓
2	Epoxymurin-A	9.91	0	3	530.88	×
3	Squamocin	8.80	3	7	622.93	×
4	Anonaine	3.25	1	3	265.31	✓
5	(+)-Reticuline	2.38	2	5	329.40	✓
6	(+)-Xylopin	3.29	1	4	295.34	✓
7	Coronin	2.29	0	5	260.25	✓
8	Gallic acid	0.59	4	5	170.12	✓
9	3-O-Caffeoylquinic acid	-0.45	6	9	354.31	×
10	Kaempferol	2.17	4	6	286.24	✓
11	Quercetin	1.68	5	7	302.24	✓
12	kaempferol 3-O-robinobioside	-0.57	9	15	594.52	×
13	Nicotiflorin	-0.57	9	15	594.52	×
14	Quercetin-3-O-glucoside	-0.36	8	12	464.38	×
15	Rutin	-1.06	10	16	610.52	×
16	(-)-Loliolide	1.84	1	3	196.25	✓
17	(+)-Isololiolide	1.84	1	3	196.25	✓

18	Asimilobine	2.90	2	3	267.33	✓
19	(-)-Coreximine	1.94	2	5	327.38	✓
20	(+)-Stepharine	1.66	1	4	297.35	✓
21	Isolaureline	3.53	0	4	309.37	✓
22	Cocclaurine	2.31	3	4	285.34	✓
23	Argentinine	3.95	1	3	295.38	✓
24	(S)-Norcorydine	2.75	2	5	327.38	✓
25	Nornuciferine	3.21	1	3	281.36	✓
26	(6S,9R)-Roseoside	-0.19	5	8	386.44	✓
27	(Z)-3-Hexenyl beta-D-glucopyranoside	-0.36	4	6	262.30	✓
28	Citroside A	-0.74	5	8	386.44	✓
29	Neochlorogenic acid	-0.45	6	9	354.31	×
30	Turpinionoside A	-0.12	6	8	390.47	×
31	Sabadelin	9.91	0	3	530.88	×
32	Annonacin A	8.41	4	7	596.89	×
33	Donhexocin	7.39	6	8	614.90	×
34	Murihexocin C	5.73	6	9	628.89	×
35	Annomontacin	8.92	4	7	624.94	×
36	Annonacin	8.41	4	7	596.89	×
37	cis-Annomontacin	8.92	4	7	624.94	×
38	Annomuricatin B	-1.10	10	18	739.83	×
39	Annonacinone	8.28	3	7	594.87	×
40	Annopentocin A	7.12	5	8	612.89	×
41	Annopentocin B	7.12	5	8	612.89	×
42	Annopentocin C	7.12	5	8	612.89	×
43	cis-Annonacin	8.41	4	7	596.89	×
44	cis-Goniothalamycin	8.41	4	7	596.89	×
45	Cohibin A	9.74	2	4	548.89	×
46	Cohibin B	9.74	2	4	548.89	×
47	Diepomuricanin A	9.73	0	4	546.88	×
48	Epomusenin A	10.05	0	3	558.93	×
49	Epomusenin B	10.05	0	3	558.93	×
50	Epoxyurin B	9.91	0	3	530.88	×
51	Gigantetrocin A	8.41	4	7	596.89	×
52	Gigantetronenin	8.71	4	7	622.93	×
53	Goniothalamycin	8.41	4	7	596.89	×
54	Isoannonacin	7.10	3	7	596.89	×
55	Isoannonacin-10-one	6.91	2	7	594.87	×
56	Javoricin	8.41	4	7	596.89	×
57	Longifolicin	9.06	3	6	580.89	×
58	Methyl nicotinate	0.53	0	3	137.14	×
59	Montecristin	9.82	2	4	574.93	×
60	Muricatetrocin A	8.41	4	7	596.89	×
61	Muricatetrocin B	8.41	4	7	596.89	×
62	Muricatin B	7.57	5	8	612.89	×
63	Muridienin 3	10.19	0	2	542.93	×
64	Murihexocin A	5.73	6	9	628.89	×

65	Murihexocin B	5.73	6	9	628.89	×
66	Neoannonin	9.05	2	6	578.88	×
67	Xylomaticin	8.92	4	7	624.94	×
68	Annocatalin	8.41	4	7	596.89	×
69	cis-Corossolone	8.99	2	6	578.88	×
70	Muricin H	9.06	3	6	580.89	×
71	Muricin I	9.23	3	6	606.93	×
72	Muricin A	8.41	4	7	596.89	×
73	Muricin B	8.41	4	7	596.89	×
74	Muricin C	8.41	4	7	596.89	×
75	Muricin D	7.49	4	7	568.84	×
76	Muricin E	7.49	4	7	568.84	×
77	Muricin F	8.01	4	7	594.87	×
78	Muricin G	8.02	4	7	594.87	×
79	Cohibin C	9.90	2	4	576.95	×
80	Cohibin D	9.90	2	4	576.95	×
81	Annoreticuïn-9-one	8.28	3	7	594.87	×
82	Chatenaytrienin 1	10.01	0	2	512.86	×
83	Chatenaytrienin 2	10.01	0	2	512.86	×
84	Chatenaytrienin 3	10.14	0	2	540.92	×
85	Chatenaytrienin 4	10.14	0	2	540.92	×
86	Rollimusin	7.62	4	8	638.93	×
87	cis-Reticulatacin-10-one	9.31	2	6	606.93	×
88	cis-Solamin	9.51	2	5	564.89	×
89	cis-Uvariamicin I	9.71	2	5	592.95	×
90	cis-Uvariamicin IV	9.71	2	5	592.95	×
91	Muricapentocin	6.88	5	8	612.89	×
92	Purpurenin	7.62	4	8	638.93	×
93	Muricatacin	4.50	1	3	284.44	✓
94	Anomuricine	3.06	2	5	329.40	✓
95	(+)-Anomurine	3.34	1	5	343.42	✓
96	Annoionol A	1.70	3	3	230.35	✓
97	Annoionoside	-0.97	7	9	406.47	×
98	Annomuricin E	7.35	5	8	612.89	×
99	Annonamine	-0.83	1	3	296.39	×
100	Arianacin	8.41	4	7	596.89	×
101	Atherosperminine	4.26	0	3	309.41	✓
102	Epomuricenin B	9.91	0	3	530.88	×
103	Corepoxylone	9.34	0	5	560.86	×
104	cis-annonacin-10-one	7.11	3	7	594.87	×
105	Blumenol C	2.48	1	2	210.32	✓
106	Gigantetrocin	8.41	4	7	596.89	×
107	Muricadienin	10.07	0	2	514.88	×
108	Muricatenol	9.30	4	6	608.95	×
109	Muridienin 3	10.19	0	2	542.93	×
110	Muridienin 4	10.19	0	2	542.93	×
111	15-Palmitoylsolamin	10.50	1	6	803.31	×
112	15-Oleoylsolamin	10.54	1	6	829.35	×

113	Robustocin	9.38	1	5	562.88	×
114	Annonaine	3.25	1	3	265.31	✓
115	Anomurine	3.34	1	5	343.42	✓
116	Casuarine	-3.17	5	6	205.21	✓
117	Coreximine	1.94	2	5	327.38	✓
118	DMJ (Deoxymannojirimycin)	-2.40	5	5	163.17	✓
119	Isoboldine	2.49	2	5	327.38	✓
120	Liriodenine	3.30	0	4	275.26	✓
121	N-methylcoclaurine	2.56	2	4	299.37	✓
122	Muricinine	2.48	3	5	313.35	✓
123	Remerine	3.50	0	3	279.34	✓
124	Reticuline	2.38	2	5	329.40	✓
125	Stepharine	1.66	1	4	297.35	✓
126	Swainsonine	-1.34	3	4	173.21	✓
127	Xylopine	3.29	1	4	295.34	✓
128	Emodin	3.01	3	5	270.24	✓
129	Chlorogenic acid	-0.45	6	9	354.31	×
130	Dicaffeoylquinic acid	1.21	7	12	516.46	×
131	Feruloylquinic acid	-0.14	5	9	368.34	✓
132	Cinnamic acid	1.91	1	2	148.16	✓
133	Apigenin-6-C-glucoside	0.52	7	10	432.38	×
134	Daidzein	2.56	2	4	254.24	✓
135	Fisetin	1.97	4	6	286.24	✓
136	Gallocatechin	1.08	6	7	306.27	×
137	Genistein	2.27	3	5	270.24	✓
138	Glycitein	2.38	2	5	284.27	✓
139	Homoorientin	0.03	8	11	448.38	×
140	Isoferulic acid	1.25	2	4	194.19	✓
141	Kaempferol 3-O-rutinoside	-0.57	9	15	594.52	×
142	Luteolin 3'7-di-O-glucoside	-1.83	10	16	610.52	×
143	Morin	1.88	5	7	302.24	✓
144	Myricetin	1.39	6	8	318.24	×
145	Quercetin 3-O-glucoside	-0.36	8	12	464.38	×
146	Quercetin 3-O-neohesperidoside	-1.06	10	16	610.52	×
147	Robinetin	1.68	5	7	302.24	✓
148	Tangeretin	3.78	0	7	372.37	✓
149	Vitexin	0.52	7	10	432.38	✓
150	Caffeic acid	0.94	3	4	180.16	✓
151	Gentisic acid	1.37	3	4	154.12	✓
152	Diepomuricanin B	9.73	0	4	546.88	×
153	Annoglaxin	6.70	4	8	610.87	×
154	Annohexocin	5.73	6	9	628.89	×
155	Annomuricin A	7.35	5	8	612.89	×
156	Corossolin	9.06	3	6	580.89	×
157	Annopentocin C	7.12	5	8	612.89	×
158	Corossolone	8.99	2	6	578.88	×

Note: ✓ = qualified, × = not qualified

TABLE S2. Betweenness centrality, closeness centrality, and degree of compounds

No	Name	Betweenness centrality	Closeness centrality	Degree
1	p-Coumaric acid	0.034892097	0.36637931	30
2	Anonaine	0.008801786	0.353577371	24
3	(+)-Reticuline	0.014299228	0.341091493	25
4	(+)-Xylopinine	0.010418774	0.342190016	27
5	Coronin	0.079710764	0.358347386	30
6	Gallic acid	0.038732665	0.30227596	29
7	Kaempferol	0.029008477	0.35475793	29
8	Quercetin	0.03578424	0.365748709	29
9	(-)-Loliolide	0.030585147	0.326923077	24
10	(+)-Isololiolide	0.030585147	0.326923077	24
11	Asimilobine	0.010551658	0.358952703	25
12	(-)-Coreximine	0.011620587	0.351239669	25
13	(+)-Stepharine	0.043923983	0.313884786	29
14	Isolaureline	0.025282923	0.346655791	26
15	Coclaurine	0.013379154	0.352990033	27
16	Argentinine	0.026437044	0.346091205	25
17	(S)-Norcorydine	0.007904255	0.346091205	25
18	Nornuciferine	0.013093161	0.346655791	26
19	(6S,9R)-Roseoside	0.047730059	0.331513261	27
20	(Z)-3-Hexenyl beta-D-glucopyranoside	0.054880088	0.334119497	27
21	Citroside A	0.035457824	0.336234177	29
22	Methyl nicotinate	0.050437472	0.361394558	29
23	Muricatacin	0.087083252	0.307971014	28
24	Anomuricine	0.016857417	0.357744108	28
25	(+)-Anomurine	0.016238306	0.353577371	28
26	Annoionol A	0.042973403	0.315281899	30
27	Annonamine	0.013165109	0.346655791	26
28	Atherosperminine	0.00966975	0.340544872	24
29	Blumenol C	0.051661084	0.352404643	29
30	Annonaine	0.009052953	0.353577371	24
31	Anomurine	0.016857417	0.357744108	28
32	Casuarine	0.045626759	0.359560068	26
33	Coreximine	0.011620587	0.351239669	25
34	DMJ (Deoxymannojirimycin)	0.033335336	0.328438949	23
35	Isoboldine	0.010323133	0.358952703	25
36	Liriodenine	0.070717324	0.328438949	27
37	N-methylcoclaurine	0.011212004	0.356543624	26
38	Muricinine	0.007986528	0.346655791	26
39	Remerine	0.005613653	0.324427481	26
40	Reticuline	0.010008757	0.351821192	23
41	Stepharine	0.043923983	0.313884786	29
42	Swainsonine	0.034960183	0.329968944	26
43	Xylopinine	0.009376627	0.342190016	27

44	Emodin	0.049208969	0.352404643	27
45	Feruloylquinic acid	0.046048729	0.347790507	29
46	Cinnamic acid	0.025251299	0.332550861	28
47	Daidzein	0.026894829	0.353577371	27
48	Genistein	0.020785486	0.352404643	25
49	Glycitein	0.022896283	0.342190016	27
50	Isoferulic acid	0.029127108	0.360780985	28
51	Morin	0.049329588	0.343851133	30
52	Robinetin	0.027601314	0.365120275	28
53	Tangeretin	0.05422869	0.349506579	27
54	Caffeic acid	0.035556314	0.341091493	27
55	Gentisic acid	0.03519748	0.301846591	28
56	Fisetin	0.06314154	0.337301587	30

TABLE S3. Betweenness centrality, closeness centrality, and degree of the target

No	Name	Betweenness centrality	Closeness centrality	Degree
1	AKR1B1	0.003419483	0.299717913	8
2	CA2	0.010064459	0.32003012	12
3	PTPN1	0.005232201	0.289115646	5
4	TLR4	0.001030997	0.281084656	3
5	CA1	0.005424507	0.306637807	10
6	MMP9	3.10E+10	0.282204515	4
7	ALOX5	0.00176057	0.294729542	5
8	CA9	0.00750802	0.306637807	9
9	HSD11B1	0	0.268308081	1
10	HCAR2	1.44E+12	0.276692708	3
11	DRD2	0.002700798	0.292297111	23
12	HTR2A	0.002702629	0.293103448	16
13	SLC6A4	0.002350435	0.285234899	9
14	HTR2B	4.56E+12	0.281456954	12
15	HTR2C	7.09E+11	0.284090909	9
16	HTR1D	5.04E+10	0.274547804	6
17	DRD3	4.22E+12	0.282579787	12
18	HTR1B	5.52E+10	0.271045918	5
19	DRD5	2.27E+11	0.278141361	8
20	DRD1	0.001685171	0.289509537	20
21	ADRA1A	7.32E+11	0.28638814	13
22	HTR1A	3.22E+12	0.280343008	9
23	SIGMAR1	0.015027855	0.309766764	15
24	SLC6A3	0.001881118	0.293508287	14
25	DRD4	3.48E+11	0.282204515	11
26	ADRA1D	1.69E+12	0.279605263	8
27	ADRA1B	1.67E+11	0.263320942	4
28	HTR7	6.23E+10	0.274193548	7
29	CCR1	0.001668414	0.272785623	2
30	PIK3CA	0	0.263975155	1

31	HSP90AA1	0	0.263975155	1
32	PTGS2	0.002155619	0.283711615	4
33	PIK3CB	0	0.263975155	1
34	NOS2	0.00400527	0.292297111	4
35	ALPL	0	0.263975155	1
36	ADORA2B	0	0.263975155	1
37	FLT1	0	0.263975155	1
38	IGF1R	4.57E+11	0.279605263	3
39	ALK	0	0.232240437	1
40	SERPINE1	0	0.232240437	1
41	CA12	0.002850776	0.299295775	8
42	CA14	3.86E+12	0.279237845	4
43	BCL2L1	0	0.232240437	1
44	CA4	0.001959702	0.29432133	6
45	NOX4	1.69E+12	0.278141361	4
46	XDH	0.002523969	0.295549374	7
47	TYR	0.002266947	0.287162162	5
48	MAOA	4.51E+12	0.284471218	5
49	ALOX5	0	0.262022195	1
50	ACHE	8.83E+09	0.267969735	2
51	ABCC1	2.80E+11	0.278505898	3
52	AVPR2	1.16E+11	0.270700637	2
53	CYP19A1	0.004454341	0.297619048	7
54	EGFR	0.002264381	0.290300546	6
55	F2	1.56E+12	0.2763329	2
56	PPARG	4.62E+09	0.246806039	2
57	ABCC9	4.62E+09	0.246806039	2
58	PLA2G1B	4.62E+09	0.246806039	2
59	AR	0.001459478	0.282579787	4
60	CYP11B1	4.62E+09	0.246806039	2
61	PGR	4.62E+09	0.246806039	2
62	PPP2CA	4.62E+09	0.246806039	2
63	HTR5A	8.92E+09	0.270356234	4
64	HTR6	4.67E+10	0.274547804	6
65	F3	1.25E+11	0.267969735	3
66	HTR1F	1.59E+10	0.23957159	2
67	MPO	1.59E+10	0.23957159	2
68	ROCK2	0	0.239032621	1
69	PIM2	1.59E+10	0.23957159	2
70	JAK3	1.59E+10	0.23957159	2
71	JAK1	1.59E+10	0.23957159	2
72	JAK2	1.59E+10	0.23957159	2
73	PRKCE	0.001420707	0.253579952	3
74	ADRA2A	0	0.257575758	1
75	ADRA2C	0	0.257575758	1
76	ADRB2	1.92E+10	0.268308081	5
77	ADRB1	1.07E+12	0.277777778	6
78	TBXA2R	8.85E+09	0.267632242	4

79	ADRB3	8.85E+09	0.267632242	4
80	HRH3	0	0.257263923	1
81	SLC6A2	1.27E+11	0.267295597	2
82	AOC3	0	0.257263923	1
83	OPRM1	5.79E+11	0.27997365	3
84	PTPRCAP	0	0.257575758	1
85	SLC5A2	1.29E+11	0.262345679	2
86	ADA	1.29E+11	0.262345679	2
87	ADORA2A	0.002348895	0.293508287	5
88	ADORA3	0.001018328	0.277053455	3
89	HDAC1	0	0.24912075	1
90	SLC5A1	0	0.24912075	1
91	SLC29A1	0	0.24912075	1
92	FUCA1	0.003199045	0.290697674	6
93	PPM1B	0	0.24912075	1
94	STAT3	0	0.250589623	1
95	IL2	1.75E+12	0.261377614	2
96	ADORA1	0.002079948	0.29432133	5
97	ADK	0	0.250589623	1
98	GAPDH	0	0.250589623	1
99	GAA	8.36E+10	0.278871391	4
100	VEGFA	0	0.250589623	1
101	EPHX2	0	0.251777251	1
102	METAP2	0	0.251777251	1
103	DDO	0	0.265625	1
104	TRPA1	0	0.265625	1
105	PIM1	3.08E+12	0.275974026	2
106	KLKB1	0	0.265625	1
107	PRKCA	0	0.235587583	1
108	PDE4A	0	0.235587583	1
109	HMGCR	0	0.235587583	1
110	PRKCG	0	0.235587583	1
111	PRKCH	0	0.235587583	1
112	PRKCD	0	0.235587583	1
113	OPRK1	0	0.235587583	1
114	TRPM8	0	0.239841986	1
115	HSD11B1	3.09E+11	0.2763329	2
116	ESR1	0.001036036	0.286002692	5
117	ESR2	0.001879392	0.292699725	6
118	CDC25A	0	0.239841986	1
119	HSD11B2	0	0.260736196	1
120	NR3C1	0	0.260736196	1
121	SERPINA6	0	0.260736196	1
122	MAPK3	0	0.260736196	1
123	MGAM	0.001445826	0.282579787	5
124	GANAB	1.47E+12	0.268987342	3
125	SI	1.47E+12	0.268987342	3

126	MANBA	5.88E+10	0.267632242	2
127	GBA2	1.47E+12	0.268987342	3
128	PLA2G2A	0	0.264632628	1
129	UGCG	0	0.247380675	1
130	GBA	1.19E+11	0.249413146	2
131	GLA	0	0.247380675	1
132	GLB1	1.19E+11	0.249413146	2
133	IDH1	0	0.247380675	1
134	GSK3B	0	0.247380675	1
135	MAPK8	0	0.247380675	1
136	MAPK10	0	0.247380675	1
137	MAPK9	0	0.247380675	1
138	CDK2	4.70E+11	0.259146341	2
139	TGFB1	0	0.247380675	1
140	ROCK2	0	0.239032621	1
141	MAN2B1	0	0.248247664	1
142	CSNK2A1	0	0.260736196	1
143	ELANE	2.60E+12	0.275974026	2
144	FNTA FNTB	0	0.260736196	1
145	MCL1	0	0.260736196	1
146	BCL2	0	0.260736196	1
147	MMP2	7.09E+10	0.272785623	2
148	AKR1B10	0	0.258201701	1
149	ALOX5	0	0.258201701	1
150	MMP12	0	0.258201701	1
151	CA5B	1.55E+10	0.271392082	2
152	ALDH2	0	0.261377614	1
153	TBXAS1	7.01E+09	0.263647643	2
154	CDK6	0	0.252375297	1
155	ALOX15	0	0.252375297	1
156	ALOX12	0	0.252375297	1
157	ARG1	0	0.252375297	1
158	CA5A	0	0.265293383	1
159	DAPK1	0	0.256024096	1
160	MPG	0	0.256024096	1
161	SLC22A12	0	0.256024096	1
162	FLT3	7.75E+09	0.274547804	2
163	OPRD1	0	0.259146341	1
164	ABCG2	0	0.259146341	1
165	CYP1B1	0	0.259146341	1
166	KIT	0	0.259146341	1
167	CA7	1.01E+12	0.266624843	2
168	CA6	1.01E+12	0.266624843	2
169	MMP1	0	0.254491018	1
170	LDHA	0	0.2319869	1
171	LDHB	0	0.2319869	1

TABLE S4. Betweenness centrality, closeness centrality, and degree of the disease

No	Name	Betweenness centrality	Closeness centrality	Degree
1	Rheumatoid arthritis	0.023199613	0.312960236	10
2	Alzheimer disease	0.031287725	0.350082372	23
3	Diabetic complication	0.083931122	0.36637931	28
4	Prostate cancer	0.001030997	0.281084656	3
5	Chronic glaucoma	0.005424507	0.306637807	10
6	Stroke	0.001111026	0.29109589	6
7	Asthma	0.047886814	0.362010221	32
8	Breast cancer	0.023479915	0.336767036	16
9	Lupus	7.23E+10	0.283711615	3
10	Lipid metabolism disorder; Cardiovascular disease	1.44E+12	0.276692708	3
11	Attention deficit hyperactivity disorder	0.010427305	0.307080925	21
12	Depression	0.001373707	0.289509537	15
13	Hyperprolactinaemia	0.003875351	0.288722826	13
14	Glioma	4.22E+12	0.282579787	12
15	Schizophrenia	0.023993075	0.323439878	25
16	Spontaneous abortion	0.001685171	0.289509537	20
17	Hypertension	0.002943421	0.29637378	9
18	Cough	0	0.254491018	1
19	Congestive heart failure	7.40E+11	0.28638814	13
20	Cognitive impairment	6.23E+10	0.274193548	7
21	Chronic obstructive pulmonary disease	0.001668414	0.272785623	2
22	Follicular lymphoma	0	0.263975155	1
23	Cerebrovascular ischaemia	0.002155619	0.283711615	4
24	Breast cancer	0.001698555	0.284471218	3
25	Endotoxic shock	0.00400527	0.292297111	4
26	Genetic disease	0	0.263975155	1
27	Paroxysmal supraventricular tachycardia	0	0.263975155	1
28	Renal cell carcinoma	0	0.263975155	1
29	Hormone deficiency	4.57E+11	0.279605263	3
30	Lung cancer	0	0.232240437	1
31	Thrombosis	0.001379314	0.288722826	5
32	Bacterial infection	0.002850776	0.299295775	8
33	Inborn error of metabolism	3.86E+12	0.279237845	4
34	Myelofibrosis	0	0.232240437	1
35	Open-angle glaucoma	0.001959702	0.29432133	6
36	Fibrosis	3.13E+12	0.279605263	4
37	Ischemia	0.002523969	0.295549374	7
38	Melanoma	0.002266947	0.287162162	5
39	Parkinson disease	4.51E+12	0.284471218	5
40	Gout	6.66E+11	0.283711615	4
41	Cushing disease	0.012433403	0.313884786	11
42	Inflammatory bowel disease	4.62E+09	0.246806039	2

43	Leishmaniasis	4.62E+09	0.246806039	2
44	Leukemia	0.003816075	0.29109589	5
45	Kidney Diseases	1.59E+10	0.23957159	2
46	Multiple myeloma	1.59E+10	0.23957159	2
47	Inflammation	1.59E+10	0.23957159	2
48	Essential thrombocythemia	1.59E+10	0.23957159	2
49	Polycythemia vera	1.59E+10	0.23957159	2
50	Anxiety disorder	0.001420707	0.253579952	3
51	Heart Failure	0	0.257575758	1
52	Myocardial Ischemia	0.006505817	0.293914246	8
53	Myocardial Infarction	8.85E+09	0.267632242	4
54	Hyperlipidemias	8.85E+09	0.267632242	4
55	Diabetic neuropathy	3.16E+12	0.275974026	2
56	Neuralgia	5.79E+11	0.27997365	3
57	Cardiovascular Diseases	0.007646779	0.306195965	8
58	Cardiovascular disease	5.17E+10	0.279605263	3
59	Haematological malignancy	1.29E+11	0.262345679	2
60	Hypotension	0.003638136	0.300565771	6
61	Ischemia	0.001018328	0.277053455	3
62	Angina pectoris	0	0.24912075	1
63	Atherosclerosis	0	0.250589623	1
64	Neoplasm Metastasis	1.75E+12	0.261377614	2
65	Intellectual Disability	0	0.250589623	1
66	Lateral sclerosis	0	0.250589623	1
67	Glycogen Storage Disease Type II	8.36E+10	0.278871391	4
68	Diabetes Mellitus, Type 2	1.49E+12	0.270700637	3
69	Ataxia	0	0.265625	1
70	Acute myeloid leukaemia	0.004646781	0.284852547	3
71	Spinal muscular atrophy	0	0.235587583	1
72	Acute bronchial asthma	0	0.235587583	1
73	Dyslipidemia	0	0.235587583	1
74	Severe Acute Respiratory Syndrome	0	0.235587583	1
75	Acute myocardial infarction	0	0.235587583	1
76	Erythema	0	0.235587583	1
77	Pain	0	0.239841986	1
78	Acne vulgaris	0.001036036	0.286002692	5
79	Cholelithiasis	0	0.239841986	1
80	Diabetes Mellitus, Type 1	0	0.260736196	1
81	Glucose Metabolism Disorders	0	0.260736196	1
82	Sucrase-isomaltase deficiency	1.47E+12	0.268987342	3
83	beta-Mannosidosis	5.88E+10	0.267632242	2
84	Autosomal Recessive	1.47E+12	0.268987342	3
85	Metabolic disorder	1.19E+11	0.249413146	2
86	Fabry disease	0	0.247380675	1

87	Gangliosidosis, GM1	1.19E+11	0.249413146	2
88	Cholangiocarcinoma	6.46E+11	0.274193548	2
89	Myotonic Dystrophy	0	0.247380675	1
90	Cell Transformation, Neoplastic	0	0.247380675	1
91	Discoid lupus erythematosus	0	0.247380675	1
92	Trigeminal Neuralgia	0	0.247380675	1
93	Thymic cancer	4.70E+11	0.259146341	2
94	Idiopathic pulmonary fibrosis	0	0.247380675	1
95	alpha-Mannosidosis	0	0.248247664	1
96	Crohn disease	2.60E+12	0.275974026	2
97	Hutchinson-Gilford progeria syndrome	0	0.260736196	1
98	Multiple Myeloma	0	0.260736196	1
99	Hepatic Fibrosis	7.09E+10	0.272785623	2
100	Neoplasm Invasiveness	0	0.258201701	1
101	Asthma	0	0.258201701	1
102	Anaplasia	0	0.249706228	1
103	Carbonic anhydrase va deficiency	1.55E+10	0.271392082	2
104	Breast Neoplasms	0	0.252375297	1
105	Hyperalgesia	0	0.252375297	1
106	Amino Acid Metabolism, Inborn Errors	0	0.252375297	1
107	Lung Neoplasms	0	0.256024096	1
108	Colonic Neoplasms	0	0.256024096	1
109	Leukemia	7.75E+09	0.274547804	2
110	Congenital chloride diarrhea	0	0.259146341	1
111	Obesity	0	0.259146341	1
112	Chemical and Drug Induced Liver Injury	1.01E+12	0.266624843	2
113	Non-Small-Cell Lung	0	0.254491018	1
114	Adenocarcinoma of Lung	0	0.2319869	1
115	Prostatic Neoplasms	0	0.2319869	1

TABLE S5. Betweenness centrality, closeness centrality, and degree of the pathway

No	Name	Betweenness centrality	Closeness centrality	Degree
1	Fructose and mannose metabolism	0.003419483	0.299717913	8
2	Oxidative phosphorylation	0.031287725	0.350082372	23
3	AGE-RAGE signaling pathway	0.02616324	0.326923077	21
4	Steroid hormone biosynthesis	0.002882211	0.290697674	4
5	Bacterial invasion of epithelial cells	0.005424507	0.306637807	10
6	Metabolism of proteins	0.042596434	0.345528455	15
7	Cytokine-cytokine receptor interaction	0.06729194	0.377442274	34
8	MAPK signaling pathway	0.09828744	0.394248609	31
9	Systemic lupus erythematosus	7.23E+10	0.283711615	3

10	cAMP signaling pathway	0.011551359	0.29637378	6
11	Regulation of actin cytoskeleton	0.00682588	0.298036466	18
12	Calcium signaling pathway	0.001373707	0.289509537	15
13	Neuroactive ligand-receptor interaction	0.040495946	0.332550861	23
14	Adrenergic signaling in cardiomyocytes	4.59E+10	0.271392082	5
15	Inflammatory mediator regulation of TRP channels	1.38E+12	0.268647282	2
16	Adrenergic signaling in cardiomyocytes	7.40E+11	0.28638814	13
17	Regulation of actin cytoskeleton	0.001428744	0.291895604	12
18	Chemokine signaling pathway	0.001668414	0.272785623	2
19	Acute myeloid leukemia	0.003714176	0.274547804	3
20	Metabolism of lipids and lipoproteins	0.002155619	0.283711615	16
21	Estrogen signaling pathway	0.006833995	0.303571429	7
22	Metabolic pathways	0.05786343	0.360169492	18
23	Calcium signaling pathway	0	0.263975155	1
24	Citrate cycle (TCA cycle)	0	0.263975155	1
25	Signaling by Type 1 Insulin-like Growth Factor 1 Receptor (IGF1R)	4.57E+11	0.279605263	3
26	Hemostasis	0	0.232240437	1
27	Nitrogen metabolism	0.002850776	0.299295775	8
28	Metabolism	3.86E+12	0.279237845	4
29	Apoptosis	5.65E+11	0.259146341	2
30	Bacterial invasion of epithelial cells	0.001959702	0.29432133	6
31	Cellular responses to stress	3.13E+12	0.279605263	4
32	Tyrosine metabolism	0.002266947	0.287162162	5
33	Interleukin-1 signaling	2.80E+11	0.278505898	3
34	AGE-RAGE signaling pathway	0	0.267969735	1
35	Cytokine-cytokine receptor interaction	4.62E+09	0.246806039	2
36	AGE-RAGE signaling pathway	4.96E+11	0.274902975	3
37	Arachidonic acid metabolism	4.62E+09	0.246806039	2
38	Pathways in cancer	0.005207229	0.29109589	5
39	Signal Transduction	0.046448729	0.351821192	19
40	Complement and coagulation cascades	4.19E+12	0.281084656	4
41	Immune System	0.005755067	0.281084656	4
42	VEGF signaling pathway	1.59E+10	0.23957159	2
43	Signaling by Interleukins	1.59E+10	0.23957159	2
44	PI3K-Akt signaling pathway	1.59E+10	0.23957159	2
45	Jak-STAT signaling pathway	0.00394087	0.278141361	3
46	cGMP-PKG signaling pathway	0	0.257575758	1
47	Signal Transduction	1.92E+10	0.268308081	5
48	Cell adhesion molecules	1.07E+12	0.277777778	6
49	Thromboxane signalling through TP receptor	8.85E+09	0.267632242	4
50	Adrenoceptors	8.85E+09	0.267632242	4
51	Beta-Alanine metabolism	0	0.257263923	1

52	Signaling by GPCR	5.79E+11	0.27997365	3
53	Metabolism	0.004396942	0.298036466	5
54	HDACs deacetylate histones	0	0.24912075	1
55	Alcoholism	0	0.24912075	1
56	Oxidative phosphorylation	0	0.250589623	1
57	Galactose metabolism	8.36E+10	0.278871391	4
58	Fluid shear stress and atherosclerosis	0	0.250589623	1
59	Alanine, aspartate and glutamate metabolism	0	0.265625	1
60	Inflammatory mediator regulation of TRP channels	0	0.265625	1
61	RNA transport	0	0.235587583	1
62	AMPK signaling pathway	0	0.235587583	1
63	Aldosterone-regulated sodium reabsorption	0	0.235587583	1
64	Vascular smooth muscle contraction	0	0.235587583	1
65	Arachidonic acid metabolism	0	0.239841986	1
66	Endocrine and other factor-regulated calcium reabsorption	0.001036036	0.286002692	5
67	Bile secretion	0	0.239841986	1
68	Lysosome	5.88E+10	0.267632242	2
69	Galactose metabolism	1.19E+11	0.249413146	2
70	Central carbon metabolism in cancer	6.05E+11	0.258830694	2
71	Cardiac conduction	0	0.247380675	1
72	Base excision repair	6.04E+11	0.263647643	2
73	Cell cycle	4.70E+11	0.259146341	2
74	Cell adhesion molecules	0	0.258201701	1
75	Nitrogen metabolism	3.59E+11	0.278141361	4
76	Signaling by Interleukins	0	0.252375297	1
77	Biosynthesis of amino acids	0	0.252375297	1
78	Bladder cancer	2.96E+12	0.270356234	2
79	Interleukin-1 signaling	0	0.256024096	1
80	Pancreatic secretion	0	0.259146341	1
81	nitrogen metabolism	0	0.259146341	1
82	Cysteine and methionine metabolism	0	0.2319869	1

TABLE S6. Betweenness centrality, closeness centrality, and degree of protein-protein interaction

No	Name	Betweenness centrality	Closeness centrality	Degree
1	GAPDH	0.072068707	0.5776173	63
2	VEGFA	0.057751109	0.5714286	60
3	MAPK3	0.067927382	0.5633803	59
4	EGFR	0.060625381	0.5633803	53
5	STAT3	0.019287654	0.5387205	51
6	HSP90AA1	0.041917216	0.5387205	49
7	PTGS2	0.044649212	0.5405405	49
8	ESR1	0.06313727	0.5387205	46

9	PPARG	0.07471732	0.5333333	43
10	MMP9	0.023815556	0.5144695	42
11	IL2	0.028488024	0.5228758	38
12	BCL2L1	0.007965098	0.5031447	37
13	TLR4	0.016017596	0.5	36
14	PIK3CA	0.008553819	0.4984424	35
15	JAK2	0.01138248	0.4938272	33
16	SLC6A4	0.074537741	0.4833837	32
17	MAPK8	0.004139932	0.4984424	32
18	NR3C1	0.054204486	0.516129	32
19	MMP2	0.003707481	0.4819277	31
20	MCL1	0.003439778	0.4651163	29
21	PGR	0.009111226	0.4819277	29
22	KIT	0.004778716	0.4790419	28
23	SLC6A3	0.050810121	0.4705882	28
24	TGFB1	0.004806835	0.4776119	27
25	IGF1R	0.001799129	0.4819277	27
26	PRKCD	0.010414657	0.4705882	26
27	AR	0.009138756	0.4719764	25
28	JAK1	0.005598122	0.4610951	25
29	SLC6A2	0.019120665	0.4113111	24
30	CDK2	0.004442882	0.4664723	23
31	GSK3B	0.003048005	0.4747774	23
32	HDAC1	0.008411824	0.4819277	23
33	SERPINE1	0.002604211	0.4678363	23
34	MPO	0.004210974	0.4519774	23
35	MAOA	0.015859067	0.4301075	22
36	PRKCA	0.0121799	0.4907975	22
37	PTPN1	0.012343099	0.4705882	22
38	JAK3	0.003405998	0.4395604	21
39	NOS2	0.016745381	0.4651163	20
40	ADRB2	0.035182575	0.495356	20
41	PPP2CA	0.015929494	0.4705882	20
42	PRKCE	0.018570687	0.4637681	20
43	CYP19A1	0.009653477	0.4584527	19
44	IDH1	0.010952491	0.4221636	18
45	MMP1	0.011389356	0.4519774	18
46	CDK6	7.34E-04	0.4407713	18
47	PIK3CB	8.35E-04	0.4244032	17
48	CA9	0.024858643	0.4432133	17
49	AKR1B1	0.036097706	0.4637681	17
50	ESR2	0.003951425	0.4545455	17
51	MAPK9	8.94E-04	0.4444444	17
52	F2	0.010114125	0.4494382	17
53	FLT3	0.001153348	0.4289544	16

54	FLT1	6.03E-04	0.4371585	16
55	DRD2	0.010907104	0.4584527	15
56	ABCG2	0.011116949	0.4347826	15
57	ALK	3.75E-04	0.4383562	15
58	MGAM	0.016665931	0.4166667	15
59	MAPK10	0.001846747	0.4188482	14
60	SI	0.0143962	0.4134367	14
61	ELANE	0.002705949	0.4092072	14
62	OPRM1	0.015860046	0.4407713	14
63	ADORA2A	0.015606885	0.4532578	13
64	TBXA2R	0.009597399	0.4188482	13
65	ALDH2	0.014480515	0.4507042	13
66	ARG1	3.58E-04	0.4278075	13
67	ALOX5	0.002813818	0.4123711	13
68	BCL2	2.92E-04	0.4145078	13
69	GBA	0.036162597	0.4395604	13
70	F3	0.001650384	0.4210526	13
71	CDC25A	9.91E-04	0.4232804	13
72	NOX4	7.39E-04	0.4278075	13
73	ADORA1	0.010136302	0.4432133	13
74	GLB1	0.005438733	0.361991	13
75	HTR1A	0.010836946	0.4071247	12
76	ACHE	0.020517538	0.4395604	12
77	PRKCG	0.003644139	0.4301075	12
78	DRD1	0.004604181	0.3773585	11
79	GLA	0.003503944	0.3603604	11
80	ADRA1B	0.006143319	0.4166667	11
81	ADK	0.002804078	0.3678161	10
82	HTR2A	0.001592668	0.3931204	10
83	LDHA	0.003444853	0.4336043	10
84	ADRA2A	0.002784239	0.3960396	10
85	ADORA2B	0.004325788	0.4166667	10
86	PLA2G1B	0.001348901	0.4102564	9
87	MMP12	1.17E-04	0.4102564	9
88	CYP1B1	4.87E-04	0.4347826	9
89	ADRA2C	0.002022301	0.3940887	9
90	MANBA	8.76E-04	0.3361345	9
91	ROCK2	2.17E-04	0.4113111	9
92	PLA2G2A	0.004410631	0.4081633	9
93	ADA	0.004992618	0.3902439	9
94	HTR5A	8.47E-04	0.3426124	9
95	HTR2C	0.002036741	0.3864734	9
96	DRD4	9.80E-04	0.3755869	9
97	SLC29A1	0.001414587	0.3478261	8

98	MAN2B1	8.66E-04	0.3354298	8
99	PIM2	2.19E-04	0.3809524	8
100	TYR	0.003554552	0.4312668	8
101	SIGMAR1	0.004047615	0.3755869	8
102	ADORA3	0.00381054	0.4324324	8
103	CCR1	5.26E-05	0.3921569	8
104	TBXAS1	0.002104146	0.4040404	8
105	UGCG	0.007703922	0.3940887	8
106	PDE4A	9.45E-04	0.383693	7
107	GANAB	0.00403321	0.334728	7
108	LDHB	0.002555428	0.4188482	7
109	AOC3	0.002362426	0.4221636	7
110	HTR1B	2.02E-04	0.3485839	7
111	HTR1D	0.001662078	0.3603604	7
112	ALOX15	3.87E-04	0.3800475	7
113	PRKCH	0.002652949	0.3931204	7
114	XDH	0.003821737	0.3883495	7
115	DRD5	0.001033359	0.3547672	7
116	PIM1	1.89E-04	0.3990025	6
117	DRD3	5.67E-05	0.3470716	6
118	ALOX12	3.81E-04	0.369515	6
119	ADRA1D	4.78E-04	0.3652968	6
120	TRPA1	0.0010905	0.3508772	6
121	HRH3	0.001580526	0.3555556	6
122	ADRA1A	4.78E-04	0.3652968	6
123	OPRD1	7.21E-04	0.3874092	6
124	GAA	9.83E-06	0.3007519	6
125	HSD11B1	9.32E-04	0.383693	6
126	FUCA1	8.09E-04	0.3333333	6
127	HTR2B	3.49E-05	0.3470716	6
128	SERPINA6	9.28E-04	0.372093	6
129	OPRK1	0.001790384	0.4266667	6
130	CYP11B1	0.001155086	0.3703704	5
131	DAPK1	6.05E-06	0.4010025	5
132	ADRB1	0.001520123	0.4081633	5
133	HSD11B2	9.75E-05	0.3547672	5
134	PPM1B	2.63E-04	0.3800475	5
135	KLKB1	0.00123536	0.3669725	5
136	SLC5A2	0.005113776	0.39801	5
137	HTR7	6.46E-04	0.3426124	5
138	CA4	0.00175775	0.3397028	4
139	CSNK2A1	3.67E-05	0.3809524	4
140	HMGCR	8.10E-04	0.3970223	4
141	GBA2	2.37E-04	0.3305785	4

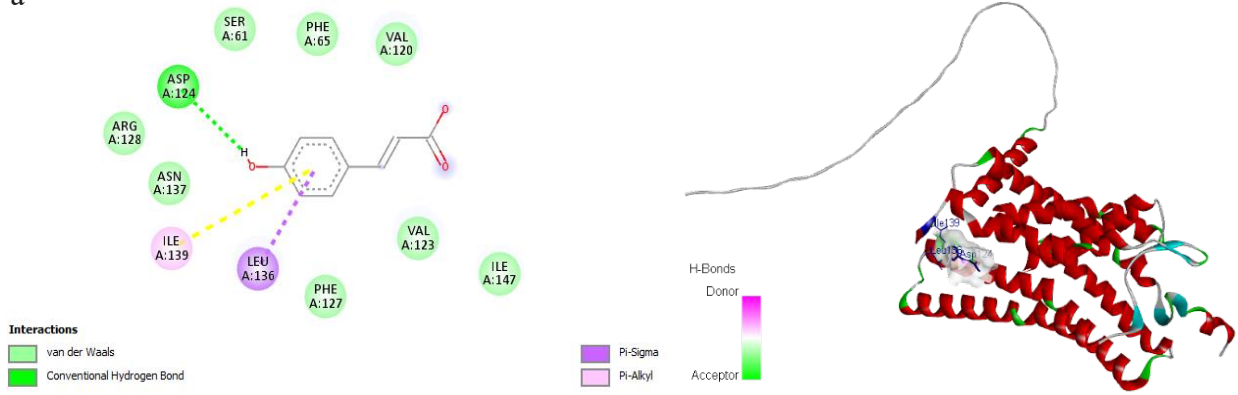
142	CA2	0.008124251	0.3539823	4
143	AKR1B10	7.10E-04	0.3501094	4
144	TRPM8	2.44E-04	0.3455724	4
145	EPHX2	5.16E-04	0.3375527	4
146	CA1	0.006835904	0.3470716	4
147	ALPL	0.004640392	0.391198	4
148	ABCC1	7.75E-04	0.3902439	4
149	CA12	0.001324298	0.3628118	4
150	AVPR2	0	0.3921569	3
151	METAP2	0	0.3940887	3
152	DDO	0	0.3319502	3
153	SLC5A1	3.58E-04	0.3782506	3
154	CA14	0	0.3131115	3
155	ADRB3	1.98E-04	0.3747073	3
156	SLC22A12	3.86E-04	0.3389831	3
157	CA7	0	0.2821869	2
158	HTR6	0	0.3292181	2
159	HTR1F	0	0.3265306	1
160	HCAR2	0	0.3485839	1
161	MPG	0	0.3508772	1
162	CA6	0	0	0
163	PTPRCAP	0	0	0
164	ABCC9	0	0	0

TABLE S7. Receptor targets, binding affinity, and amino acid residues on molecular docking analysis. Bold letters indicate hydrogen bonds formed between the ligand–receptors

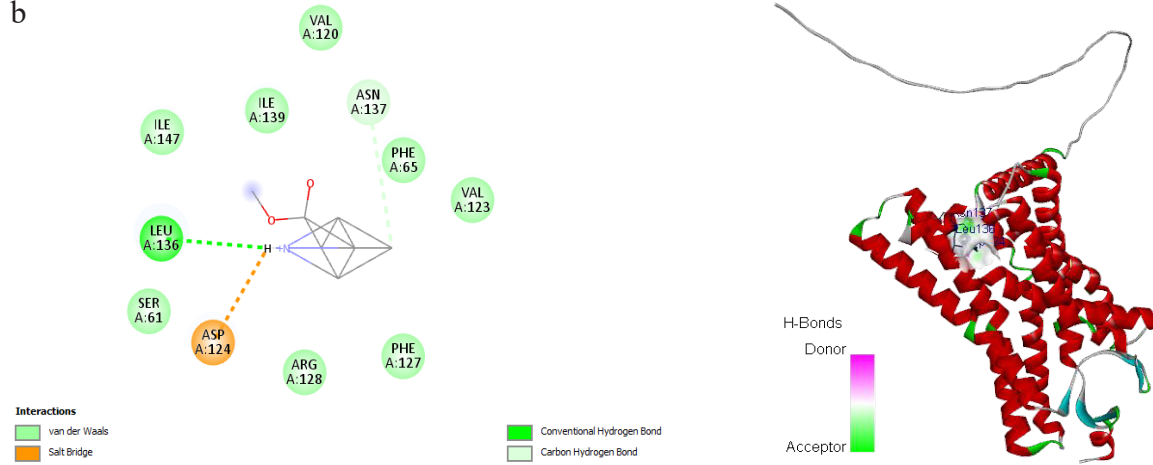
No	Ligand	Target	Binding affinity (kcal mol ⁻¹)	Amino acid residues
1	p-Coumaric acid	HCAR2	-6,2	Ser61, Phe65, Val120, Val120, Asp124 , Phe127, Arg128, Asn137, Leu136, Ile139, Ile147
2	Methyl nicotinate	HCAR2	-5,4	Ser61, Phe65, Val120, Asn124, Phe127, Arg128, Leu136 , Ile139, Asn137, Ile147
3	Cinnamic acid	HCAR2	-6,1	Ser61 , Phe65, Val120, Val123, Asp124 , Phe127, Arg128, Leu136, Asn137, Ile139, Ile147
4	Muricatacin	HMGCR	-5,9	Ala564, Tyr479, Arg495 , Lys480, Gly542, Asn567, Arg568, Glu528, Val530 , Met523, Gly524, Asn529
5	Coronin	PTGS2	-7,5	His39, Cys41, Arg44, Gly45, Val46, Cys47 , Tyr130 , Gly135, Lys137, Leu152, Pro153, Gln461, Glu465, Arg469
6	(-)-Loliolide	PTGS2	-7,0	Tyr348, Val349, Leu352, Val523, Ala527
7	(+)-Isololiolide	PTGS2	-5,8	His207, Phe210, Thr212 , His214, Asn382, Tyr385, His386, His388, Val447, Gln454
8	Coclaurine	ADRB3	-8,9	Tyr59 , Ala60, Met101, Thr102, Ser147, Arg150 , Asn230, Gln236, Thr274, Ser275, Arg314, Ser316, Cys317, Leu318 , Trp332

9	Anomuricine	ADRB3	-8,0	Tyr59 , Ala60, Met61, Met101 , Thr102, Ser147, Met188, Ser189, Asn230, Ala231, Lys233, Gln236, Thr274, Ser275, Val276, Arg314, Ser316, Cys317, Leu318
10	(+)-Anomurine	ADRB3	-6,9	Ala60, Met61, His62, Trp63, Arg150, Ser191 , Leu192, Pro194, Leu19, Phe234, Cys233, Val276, Pro236, Val276, Ser277, Phe278, Cys317, Leu318, Gly319, Val320
11	Anomurine	ADRB3	-7,6	Ala48, Asp49, Asn50, Ser51, Asp5, Asn35, Gly52, Cys107, Trp47, Asp109, Phe108, Thr111, Val110, Arg231, Asp229, Glu258, Arg255 , Asn261, Ala259, Ser265, Leu262,
12	Coclaurine	ADRB2	-7,9	Leu1013, Lys1060, Ile1050, Glu1064, Ala1063, Ile1029, Asp1061, Glu1062, Lys1065, Leu1066, Phe1067, Asn1068
13	Anomuricine	ADRB2	-7,1	Glu1011, Tyr1018, Asp1020 , Glu1022 , Tyr1024, Leu1032, Gly1030, Asp1070, Thr1026, His1031, Phe1104, Gln1105
14	(+)-Anomurine	ADRB2	-7,1	Glu1011, Asp1020 , Glu1022, Leu1032, Gln1105 , Arg1145
15	Anomurine	ADRB2	-7,0	Glu1011, Tyr1018, Asp1020 , Thr1026, Gly1030 , Leu1032, Glu1022, Tyr1024, His1031, Asp1070, Phe1104, Gln1105
16	N-methylcoclaurine	ADRB2	-7,9	Glu1011, Asp1020, Glu1022, Thr1026, Gly1030 , His1031, Leu1032, Asp1070, Val1071, Ala1074, Phe1104, Gln1105, Gly1107
17	(Z)-3-Hexenyl beta-D-glucopyranoside	VEGFA	-6,0	Phe36, Leu66
18	Kaempferol	XDH	-8,3	Leu228, Thr229, Cys285 , Arg288, Ile326 , Met329, Leu330, Leu333
19	Quercetin	XDH	-9,3	Phe226, Cys285, Gln286, Leu228, Arg288, Ser289 , Ala292, Glu295, Ile326, Tyr327, Met329, Leu330, Leu340, Leu333, Ile341, Phe363, Met364
20	Emodin	XDH	-7,1	Tyr222, Phe226, Pro227, Leu228, Thr229, Cys285, Arg288, Glu295, Leu330, Leu333, Ile326, Met329 , Ser332, Ile341, Ser342
21	Fisetin	XDH	-8,8	Tyr222, Phe226, Pro227, Leu228, Thr229, Cys285, Arg288, Glu295, Ile326 , Met329 , Leu330, Ser332, Leu333, Ile341, Ser342
22	Glycitein	XDH	-8,4	Tyr222, Phe226, Leu228 , Thr229, Lys230, Tyr222, Cys285, Ser289, Arg288, Ala292, Glu295, Met329, Leu330, Ser332, Leu333
23	Morin	XDH	-8,6	Tyr222, Phe226, Leu228 , Thr229, Cys285, Arg288, Ser289, Ala292, Glu295, Ile326, Met329, Leu330, Ser332, Leu333, Leu340
24	Robinetin	XDH	-8,8	Tyr222, Phe226, Pro227, Leu228 , Thr229, Cys285, Arg288, Glu295 , Ile326 , Met329, Leu330, Ser332, Leu333, Ile341, Ser342, Glu343

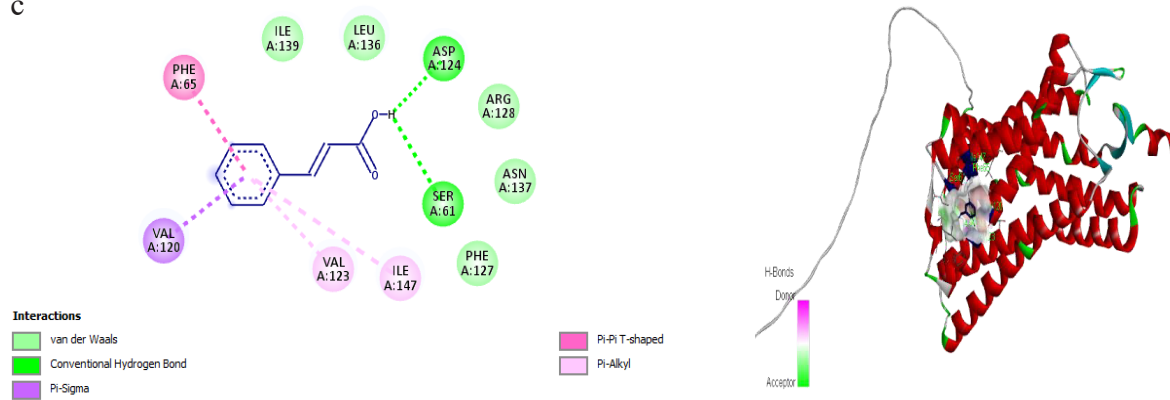
a



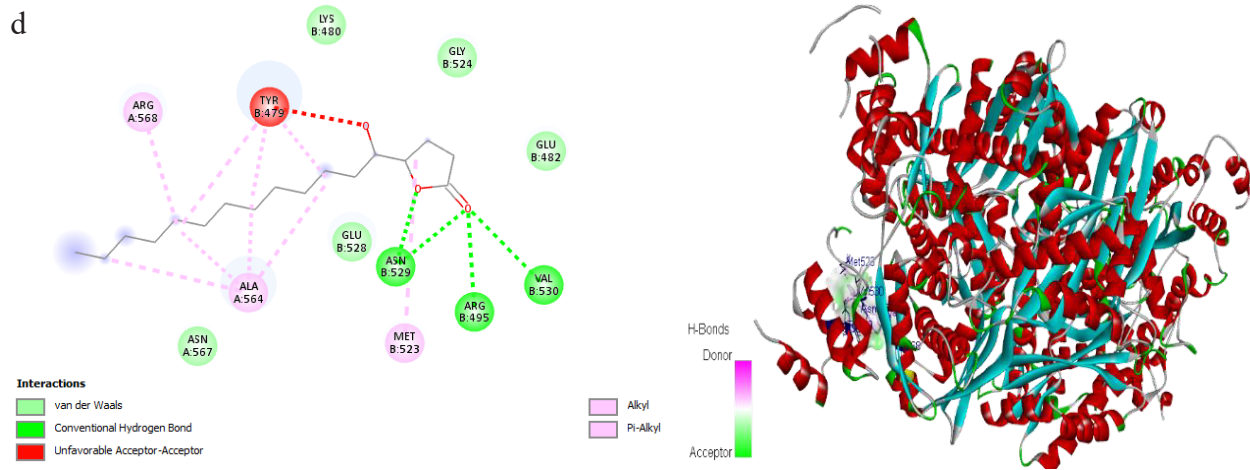
b



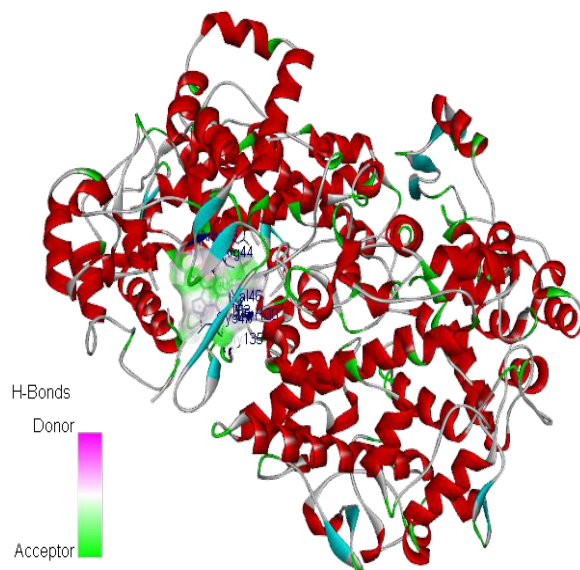
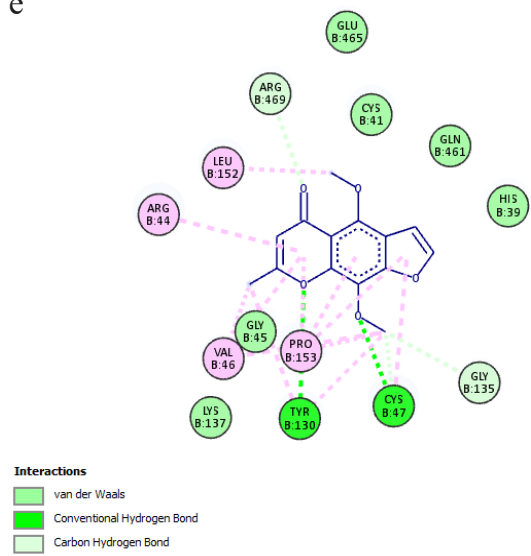
c



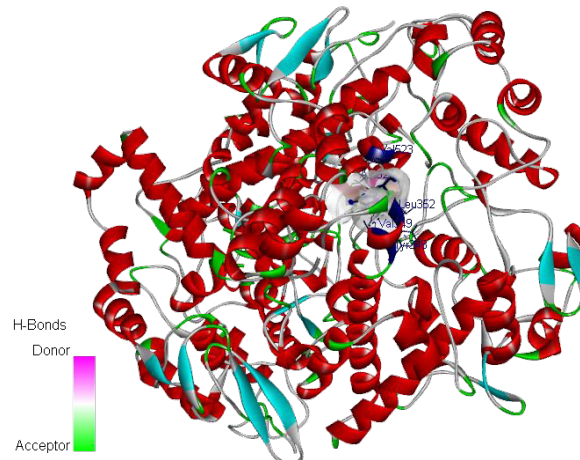
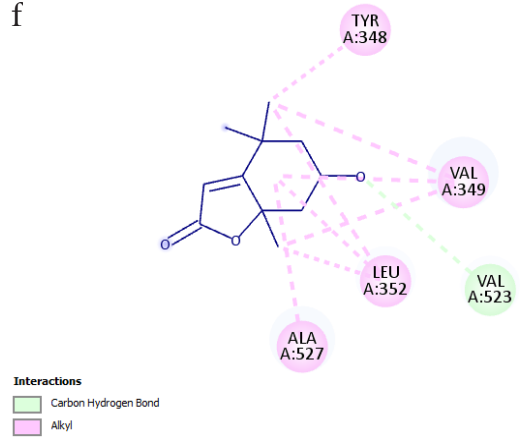
d



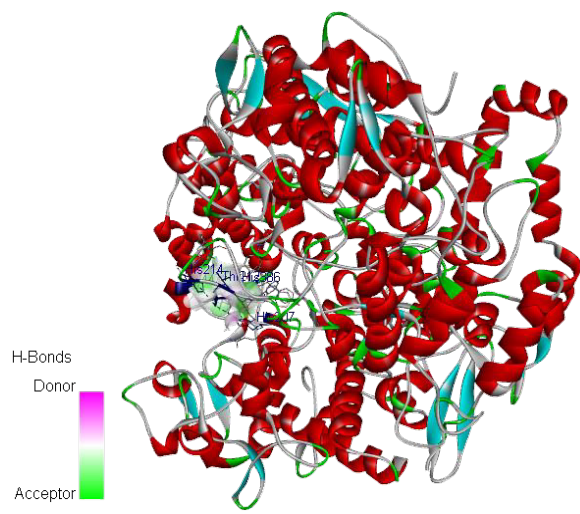
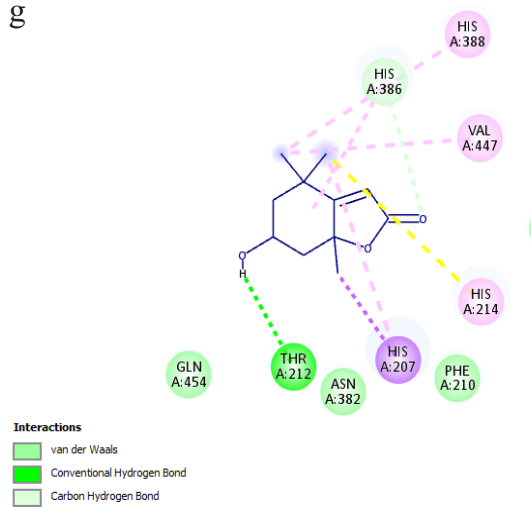
e



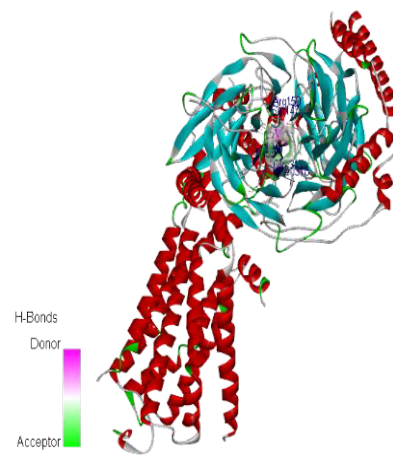
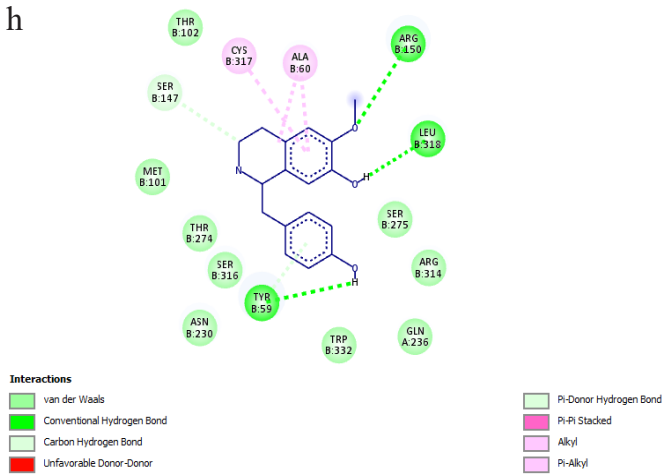
f



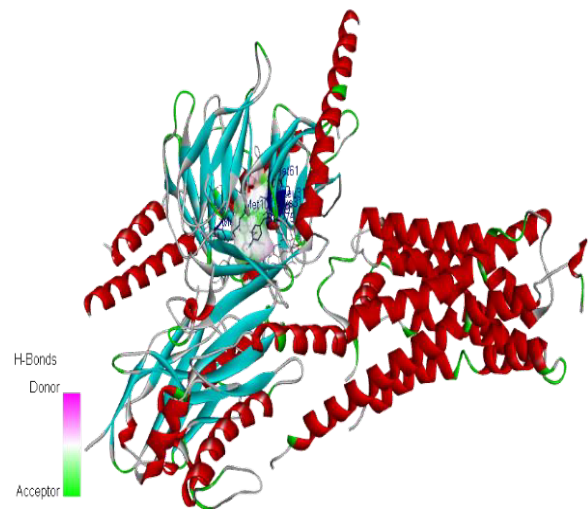
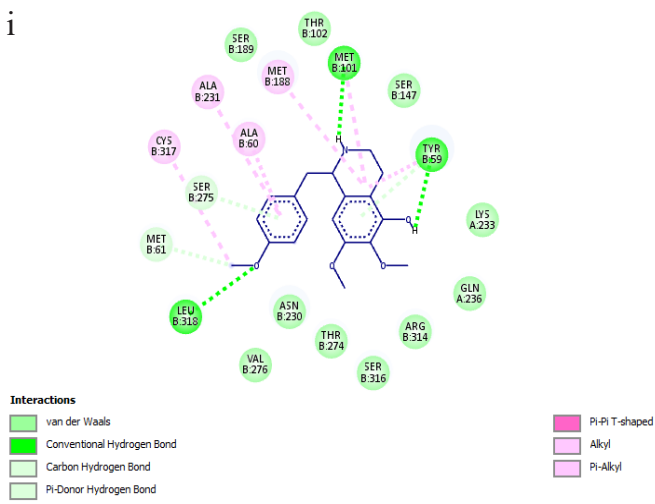
g



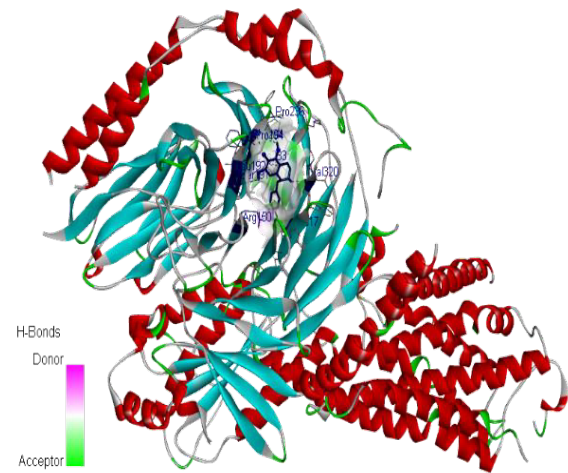
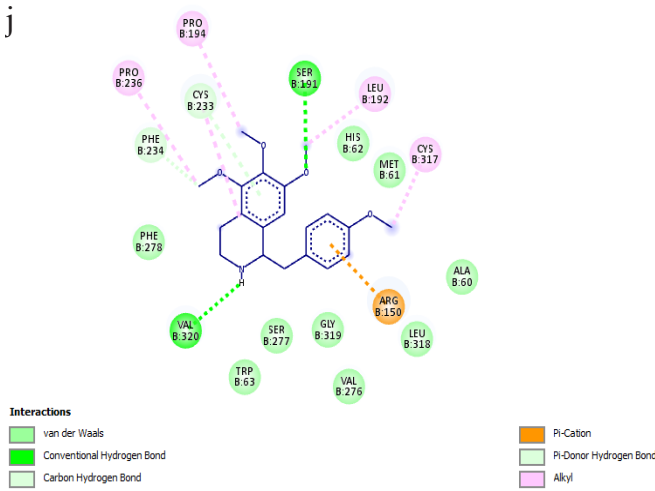
h



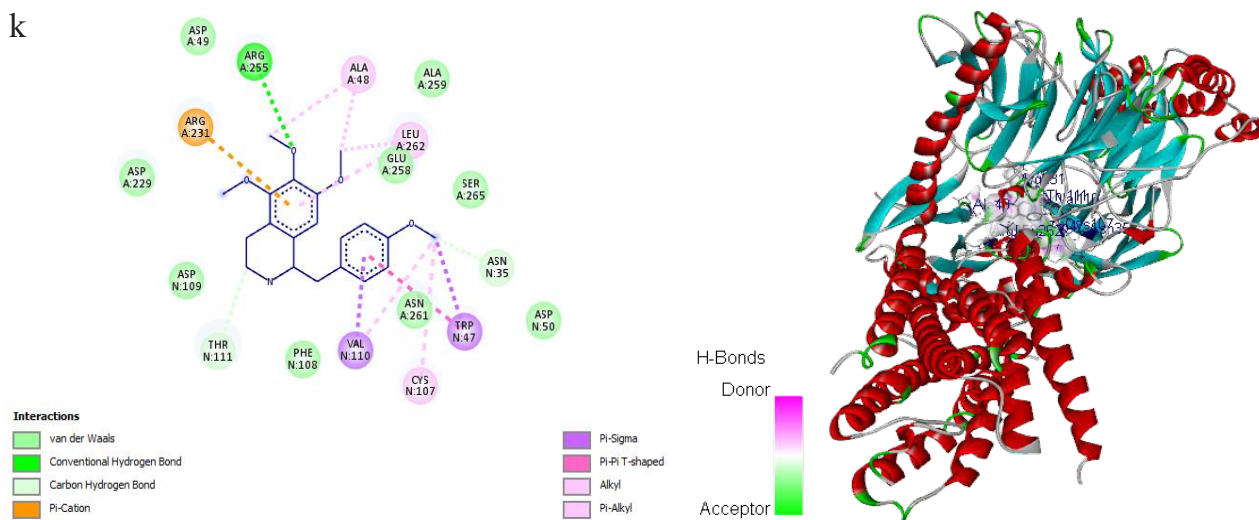
i



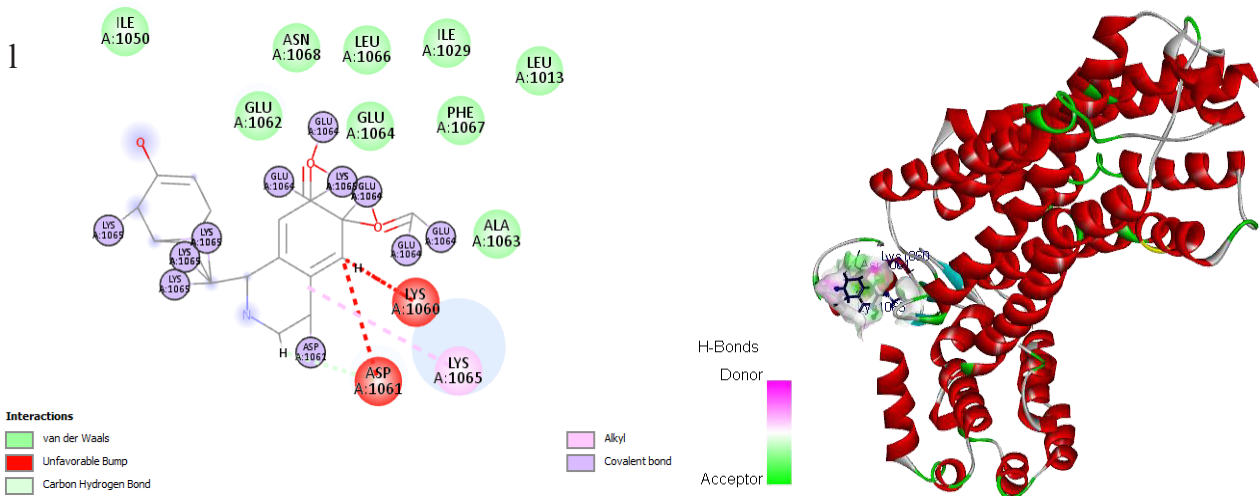
j



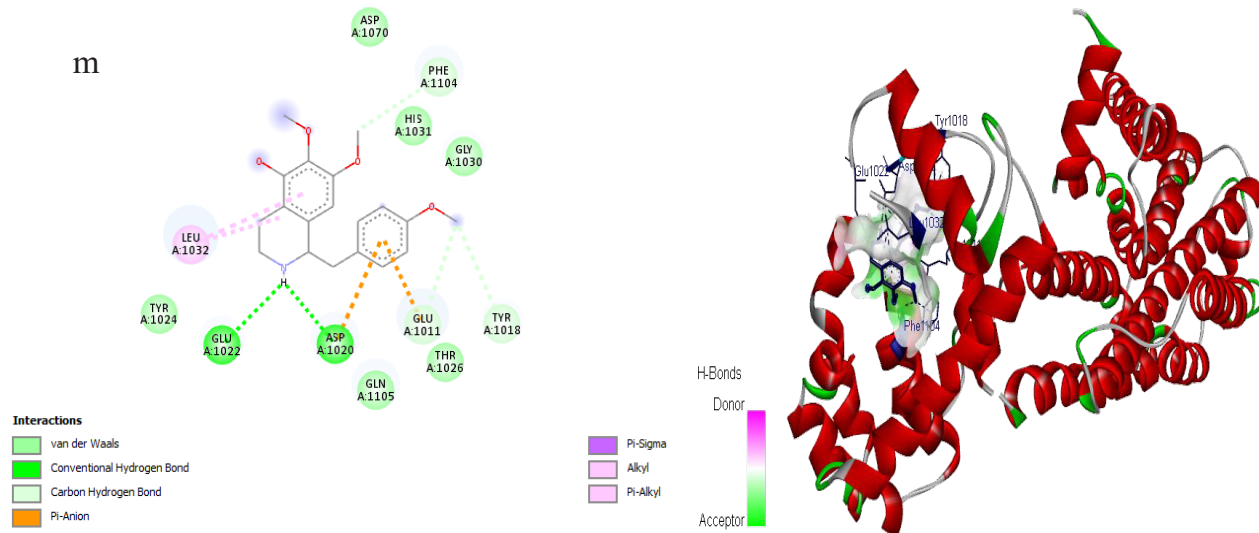
k



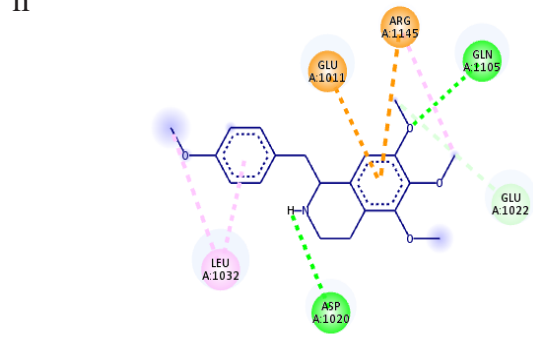
l



m



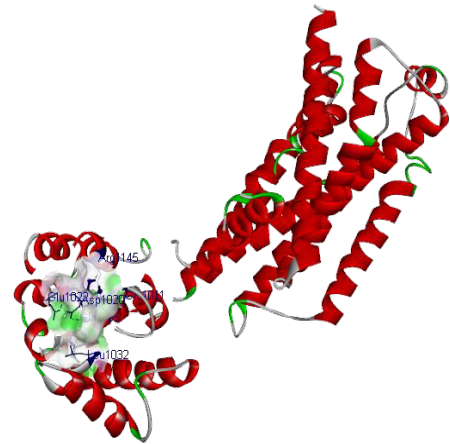
n



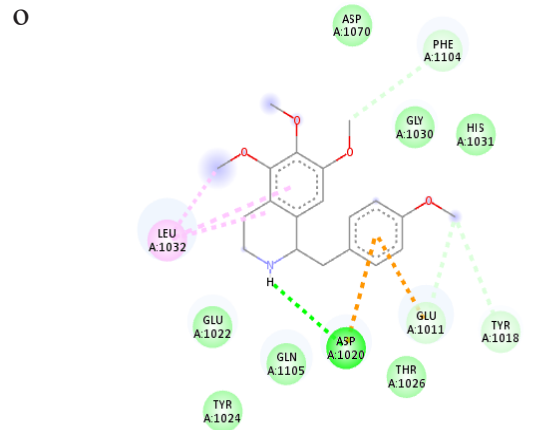
Interactions
 Conventional Hydrogen Bond
 Carbon Hydrogen Bond
 Pi-Cation

Pi-Anion
 Alkyl
 Pi-Alkyl

H-Bonds
 Donor
 Acceptor



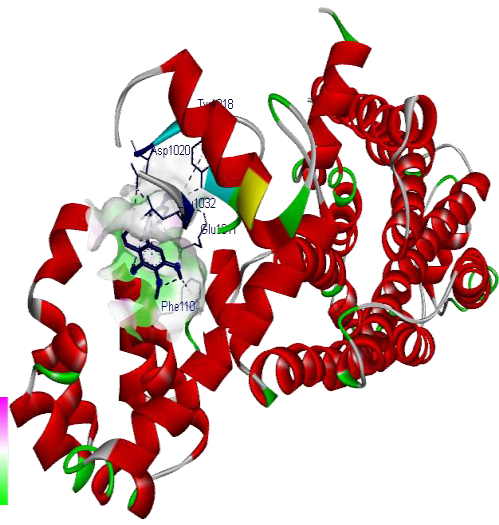
o



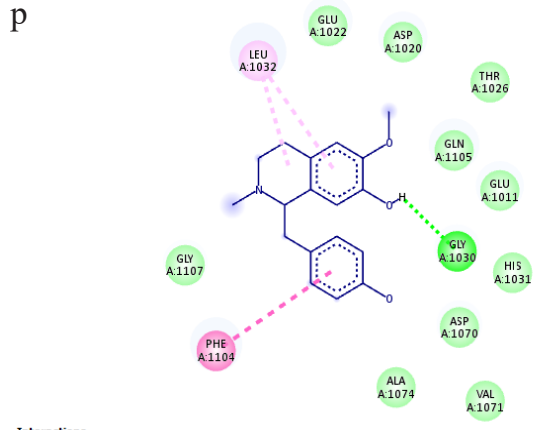
Interactions
 van der Waals
 Conventional Hydrogen Bond
 Carbon Hydrogen Bond

Pi-Anion
 Alkyl
 Pi-Alkyl

H-Bonds
 Donor
 Acceptor



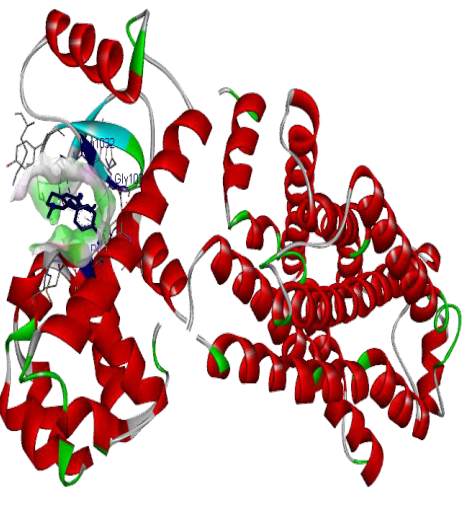
p



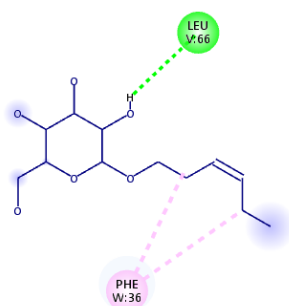
Interactions
 van der Waals
 Conventional Hydrogen Bond
 Pi-Pi T-shaped

Alkyl
 Pi-Alkyl

H-Bonds
 Donor
 Acceptor

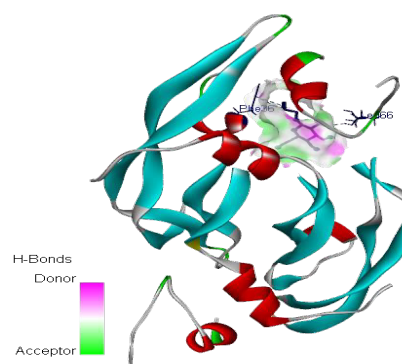


q

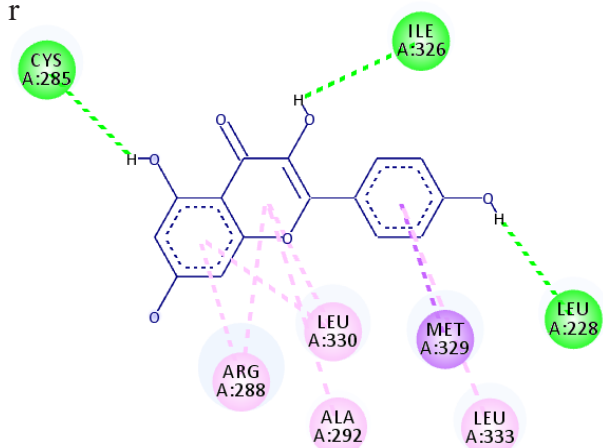
**Interactions**

Conventional Hydrogen Bond

Pi-Alkyl



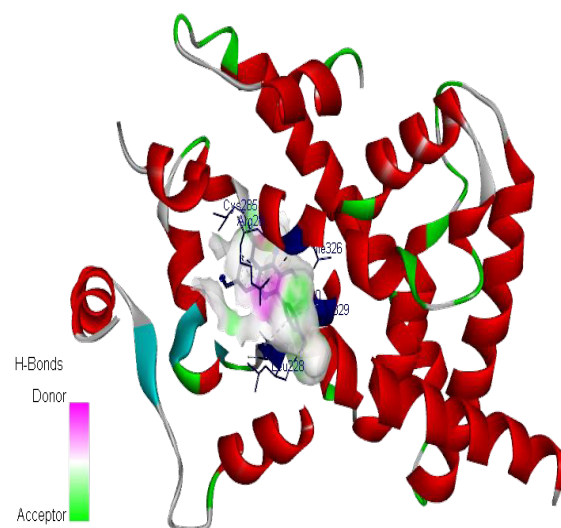
r

**Interactions**

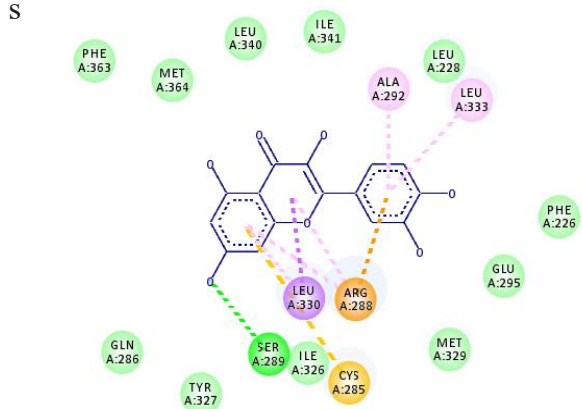
Conventional Hydrogen Bond

Pi-Sigma

Pi-Alkyl



s

**Interactions**

van der Waals

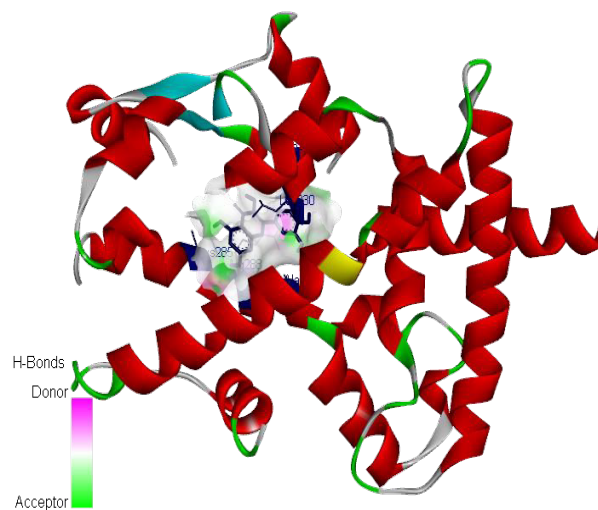
Conventional Hydrogen Bond

Pi-Cation

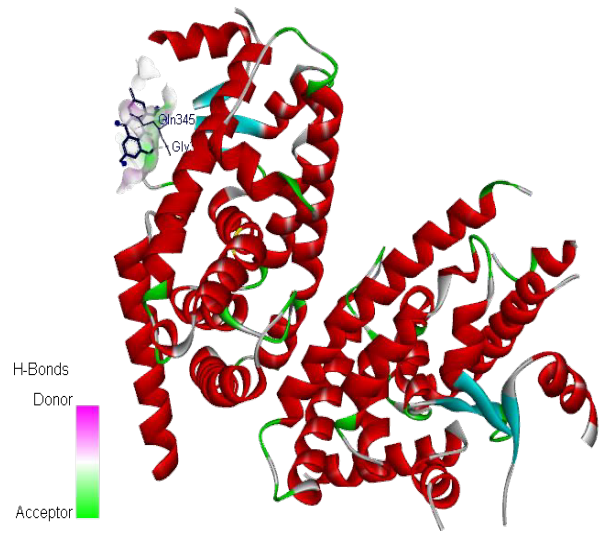
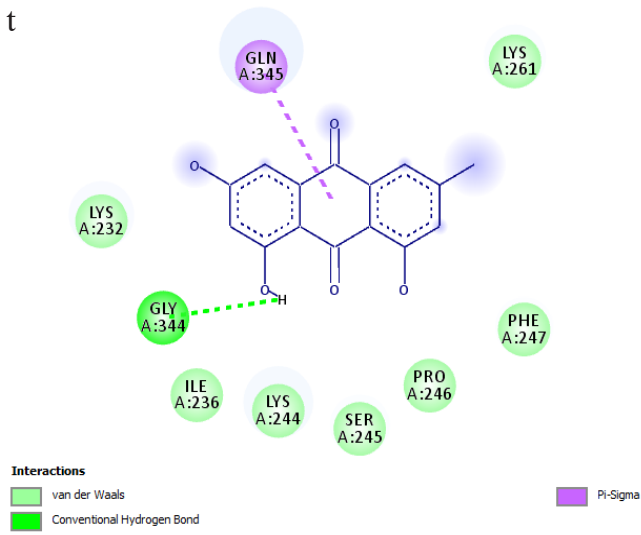
Pi-Sigma

Pi-Sulfur

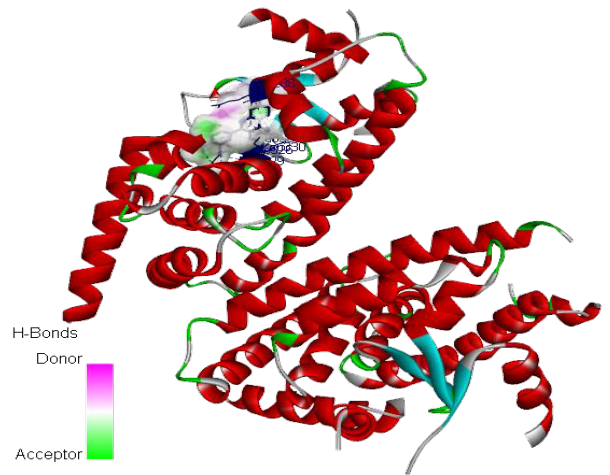
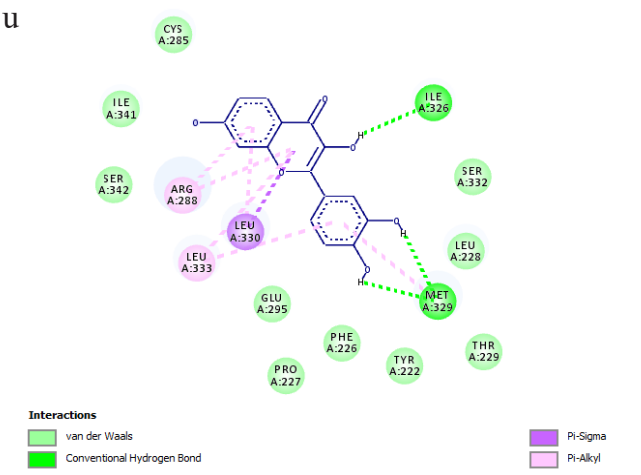
Pi-Alkyl



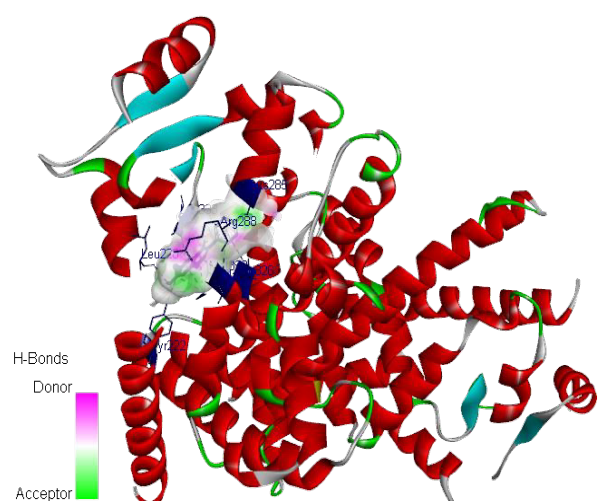
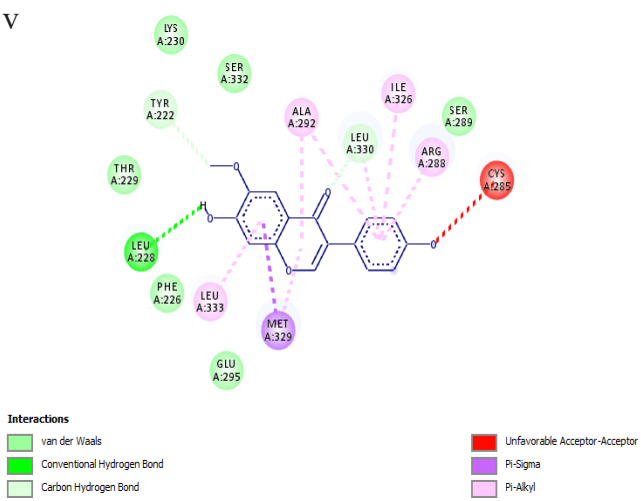
t



u



v



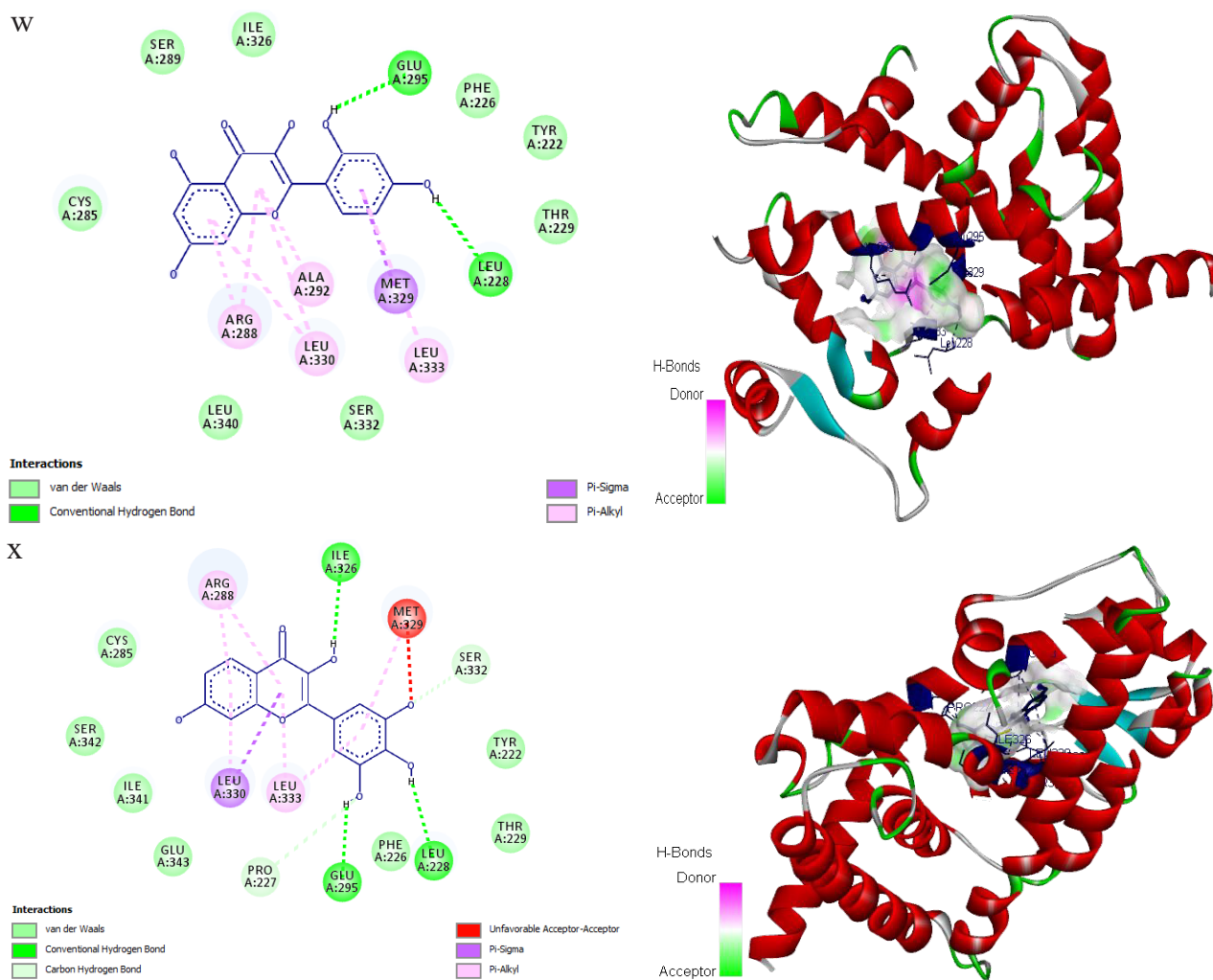


FIGURE S2. Docking compounds in 2D and 3D. Compounds of p-Coumaric acid (a), methyl nicotinate (b), and cinnamic acid (c) with HCAR2; muricatacin (d) with HMGCR; coronin (e), (-)-loliolide (f), and (+)-isololiolide (g) with PTGS2; coclaurine (h), anomuricine (i), (+)-anomurine (j), and anomurine (k) with ADRB3; coclaurine (l), anomuricine (m), (+)-anomurine (n), anomurine (o), and N-methylcoclaurine (p) with ADRB2; (Z)-3-Hexenyl beta-D-glucopyranoside (q) with VEGFA; chaempherol (r), quercetin (s), emodin (t), fisetin (u), glycitein (v), morin (w), and robinetin (x) with XDH